



IX International Course of Massive Data Analysis FOR GENOMICS



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Conclusions

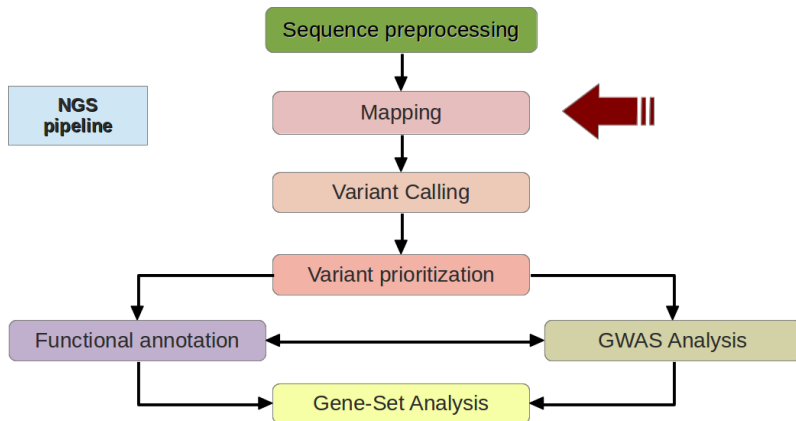
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Where are we?



Why QC on mapped sequences

Acknowledgment: Fernando García Alcalde

- ▶ The reads **may look OK** in QC analyses of **raw reads** but some **issues** only show up **once the reads are aligned**: low coverage, homopolymer biases, experimental artifacts, etc.
- ▶ These unwanted biases can be introduced by the selected:
 - ▶ Sample extraction process
 - ▶ Sequencing technology
 - ▶ Sample preparation protocol
 - ▶ Mapping algorithm

Why QC on mapped sequences

- ▶ SAM/BAM files usually contain information from tens to hundreds of millions of reads
- ▶ The **systematic detection** of such biases is a **non-trivial** task that is **crucial** to to drive appropriate downstream analyses.
- ▶ Look for big biases that really affect the analysis
- ▶ Difficult to provide guidelines: general trends

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Features

- ▶ Facilitates the identification of sequencing error biases that may disturb the mapping process
- ▶ Provides a concise html page with statistics that highlight problems in the data processing:
 - ▶ Reads with an excessive proportion of mapping errors
 - ▶ Reads containing contaminants
 - ▶ Reads representing novel splice junctions/genomic regions
 - ▶ ...
- ▶ Easy-to-use command-line tool freely downloadable at:

<http://samstat.sourceforge.net>

Timo Lassmann, Yoshihide Hayashizaki, and Carsten O. Daub. SAMStat: monitoring biases in next generation sequencing data *Bioinformatics* (2011) 27(1): 130-131

Running SAMStat

- ▶ Input: a BAM/SAM file (other sequence files are also accepted such as fasta or fastq)
- ▶ Output: an html report

Run SAMStat with a .bam example

```
samstat /home/biouser/mda13/mqc-igv/test1.bam
```

- ▶ The html report will be saved at `/home/biouser/mda13/mqc-igv/test1.bam.html`. Use a web browser (e.g. Firefox) to open it

Concepts

- ▶ Mapping quality: an integer in $[0,254]$ representing $-10 \cdot \log_{10} P(\text{mapping error})$
- ▶ Calculated as a function of the quality of the read, and a score that indicates how well the read is aligned
- ▶ Algorithm-specific
- ▶ The higher it is, the better the alignment.
(MAPQ = 30 \implies 0.001 error rate)
- ▶ 255 indicates that the mapping quality is not available.

Number of aligned reads and mapping quality

- ▶ Proportion of reads mapped in each mapping quality range.
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- ▶ **Why?**

Mean base quality

- ▶ Mean quality per read base in each mapping quality range
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Error profiles

- ▶ Number of mismatches at each read position, segregated by the nucleotide causing the mismatch
- ▶ Should be more or less stable across read positions
- ▶ More errors are expected at the end of the reads since base qualities tend to be lower at that positions
- ▶ Nucleotide peaks at different positions may indicate experimental artifacts that disturb read mapping

Over-represented di-nucleotides

- ▶ Over-representation scores for each possible di-nucleotide at each read position.
- ▶ Significant scores ($p\text{-value} \leq 1e-100$) appear in bold
- ▶ Over-represented di-nucleotides may indicate experimental artifacts that disturb read mapping

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Error distribution

- ▶ Distribution of the number of errors (mismatches and indels) per read, segregated by mapping quality ranges
- ▶ No more than ~ 2 mismatches should be allowed for short ($\sim 75b$) reads

Nucleotide composition

- ▶ Number of As,Cs,Gs and Ts appearing at each read position and segregated by mapping quality
- ▶ The counts and proportions should be almost invariant accross read positions

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Length distribution

- ▶ Distribution of the number of bases per read

Top 5 over-represented 2-mers

- ▶ Summary of the “Over-represented di-nucleotides”, including the top-5 2-mers in each position

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Top 20 over-represented 10-mers

- ▶ The 20 most significant 10-mers per quality level

Hands-on

- ▶ Run SAMstat on
`/home/biouser/mda13/mqc-igv/test2.bam` and
`/home/biouser/mda13/mqc-igv/test3.bam`

- ▶ Interpret the results

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Aim

Provide an overall view of the data that helps to detect biases in the sequencing and/or mapping of the data

Run QualiMap

qualimap

- ▶ BAM file needs to be sorted: `samtools sort <filename> <fileout>`
- ▶ File → New analysis → BAM/SAM file →
`/home/biouser/mda13/mqc-igv/HG00096.chrom20.bam`

García-Alcalde, et al. Qualimap: evaluating next generation sequencing alignment data. *Bioinformatics*(2012) 28 (20): 2678-2679

Features

- ▶ Fast analysis across the reference of genome coverage and nucleotide distribution
- ▶ Easy to interpret summary of the main properties of the alignment data
- ▶ Analysis of the reads mapped inside/outside of the regions provided in GFF format
- ▶ Insert size mean and median value calculation and plotting statistical distribution
- ▶ Analysis of the adequacy of the sequencing depth in RNA-seq experiments
- ▶ Clustering of epigenomic profiles

- ▶ Open the online help
- ▶ Go through the examples
- ▶ Run qualimap over `/home/biouser/mda13/mqc-igv/igv1.bam` (with and without reference annotation)
<http://reports.bioinfomgp.org/external-downloads/chr21.gtf>)
- ▶ Run qualimap in the previous data (with and without reference annotation <http://reports.bioinfomgp.org/external-downloads/refseqgenes.gtf>)
- ▶ Have a look at <http://reports.bioinfomgp.org/external-downloads/fullbam/qualimapReport.html>
- ▶ Drive conclusions from what you get
- ▶ BONUS: Run qualimap via de command line

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- ▶ One should always perform QC on the mapped data
- ▶ The correct interpretation of the QC output may save a lot of time (and money) on downstream analyses
- ▶ The expected results are experiment-specific \implies Learn from experience