



# MDA VII

**Joaquín Dopazo**

**Department of Bioinformatics and Genomics,  
Centro de Investigación Príncipe Felipe (CIPF),  
Functional Genomics Node, (INB), and  
Bioinformatics Group (CIBERER)  
Valencia, Spain.**

**<http://www.babelomics.org>  
<http://bioinfo.cipf.es>**

*Valencia, 21 March 2011*

# Who we are

**The Bioinformatics and Genomics Department at the Centro de Investigación Príncipe Felipe (CIPF), Valencia, Spain, and...**

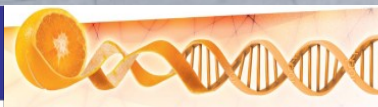


PRINCIPE FELIPE  
CENTRO DE INVESTIGACION



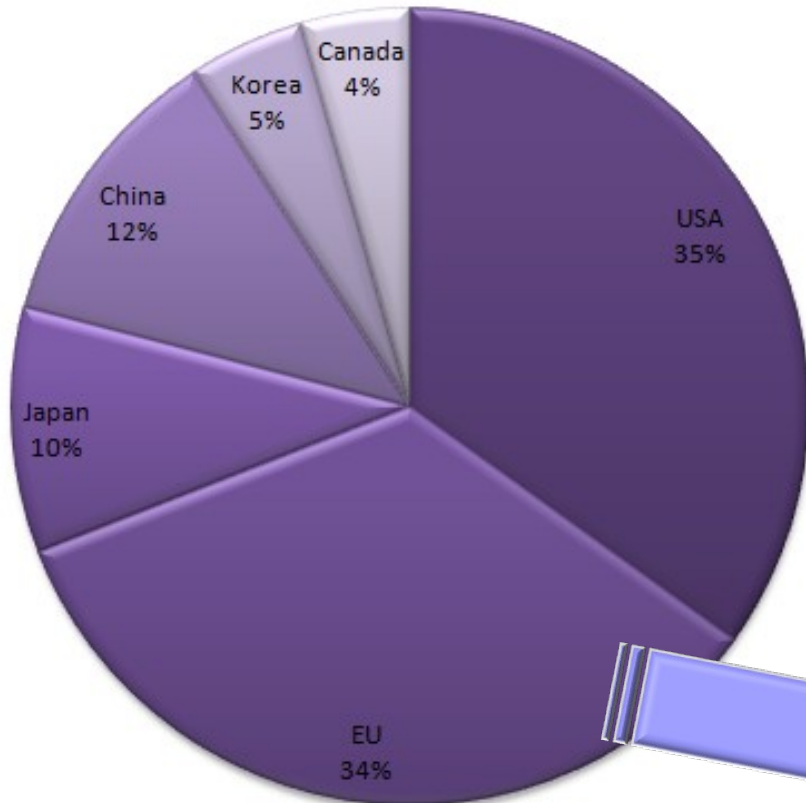
**...the INB, National Institute of Bioinformatics (Functional Genomics Node) and the CIBERER Network of Centers for Rare Diseases, and...  
...the Medical Genome Project (Sevilla)**

*ciberer*



# Some bibliographic data

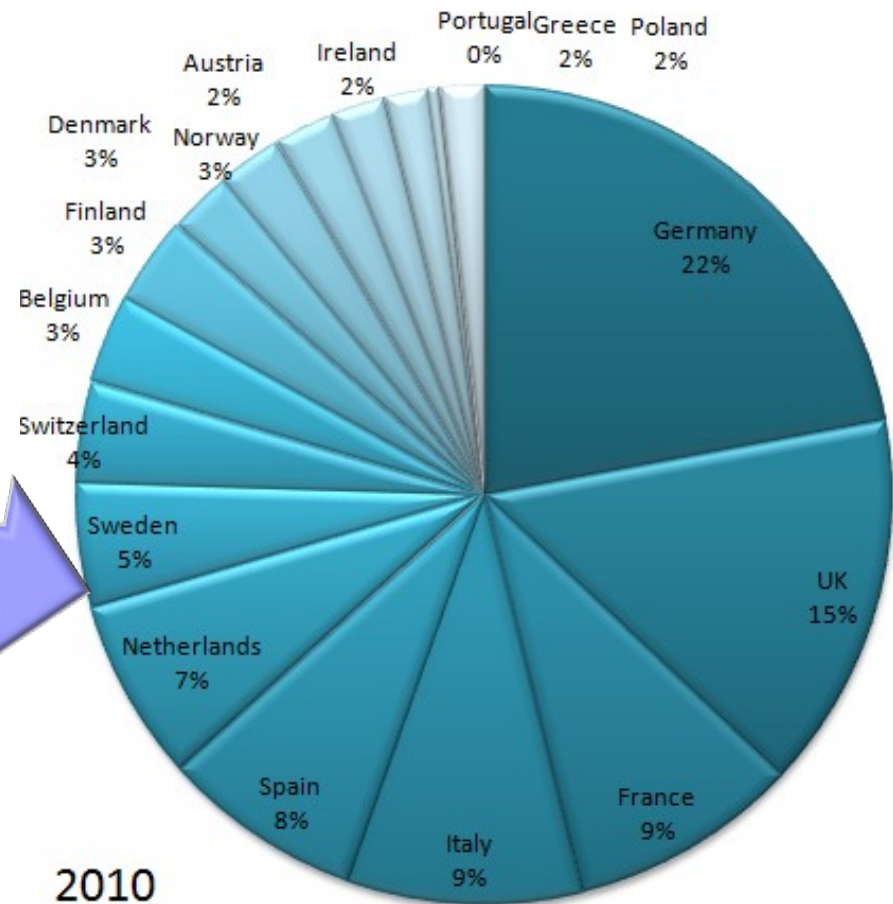
## Microarray publications



2010 Worldwide

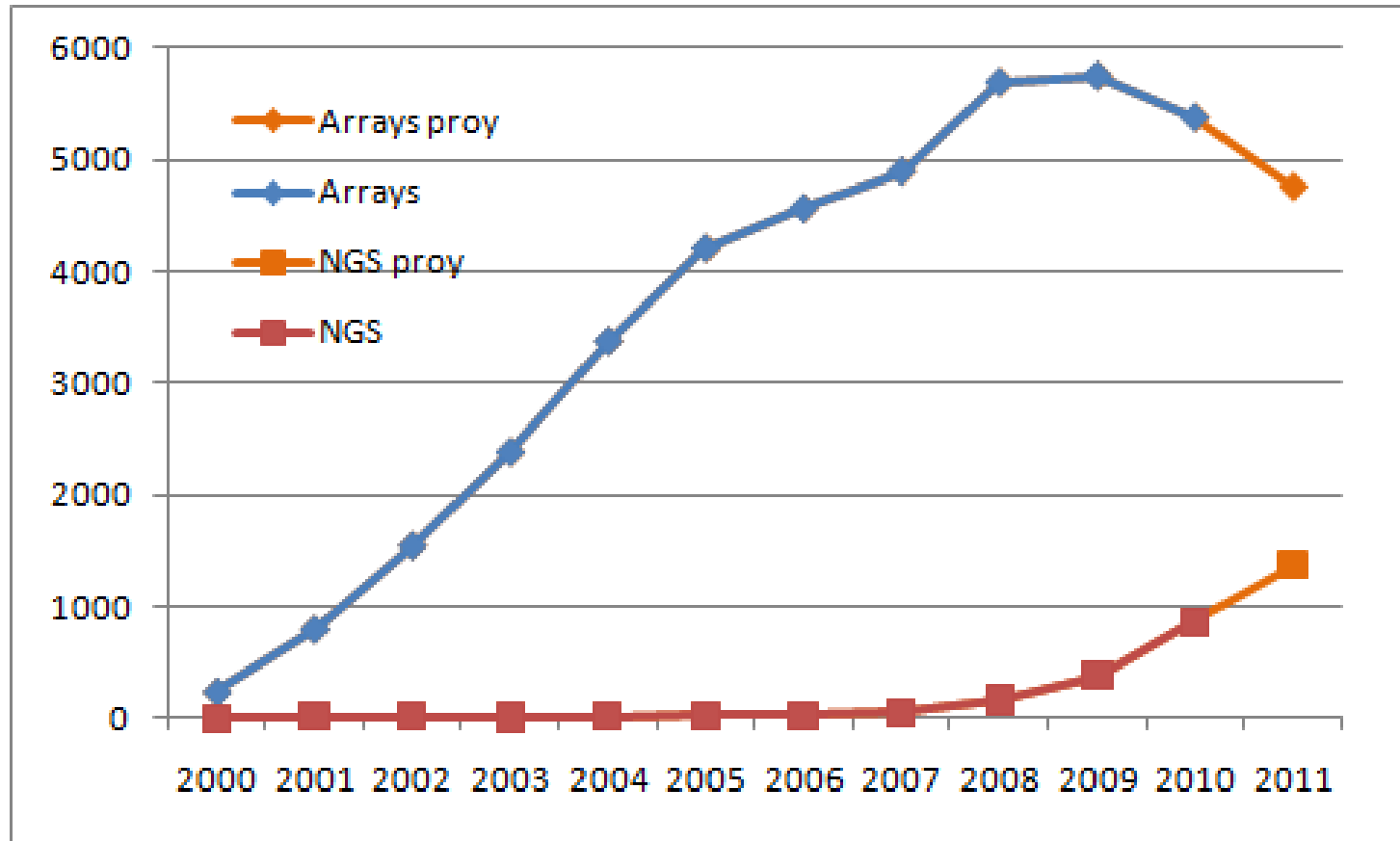
**Source Pubmed. Query:**  
**2009[Entrez Date] AND**  
**country[Affiliation]AND**  
**microarray[Title/Abstract]**

2010 Europe



2010

# Evolution of the papers published in microarray and next gen technologies

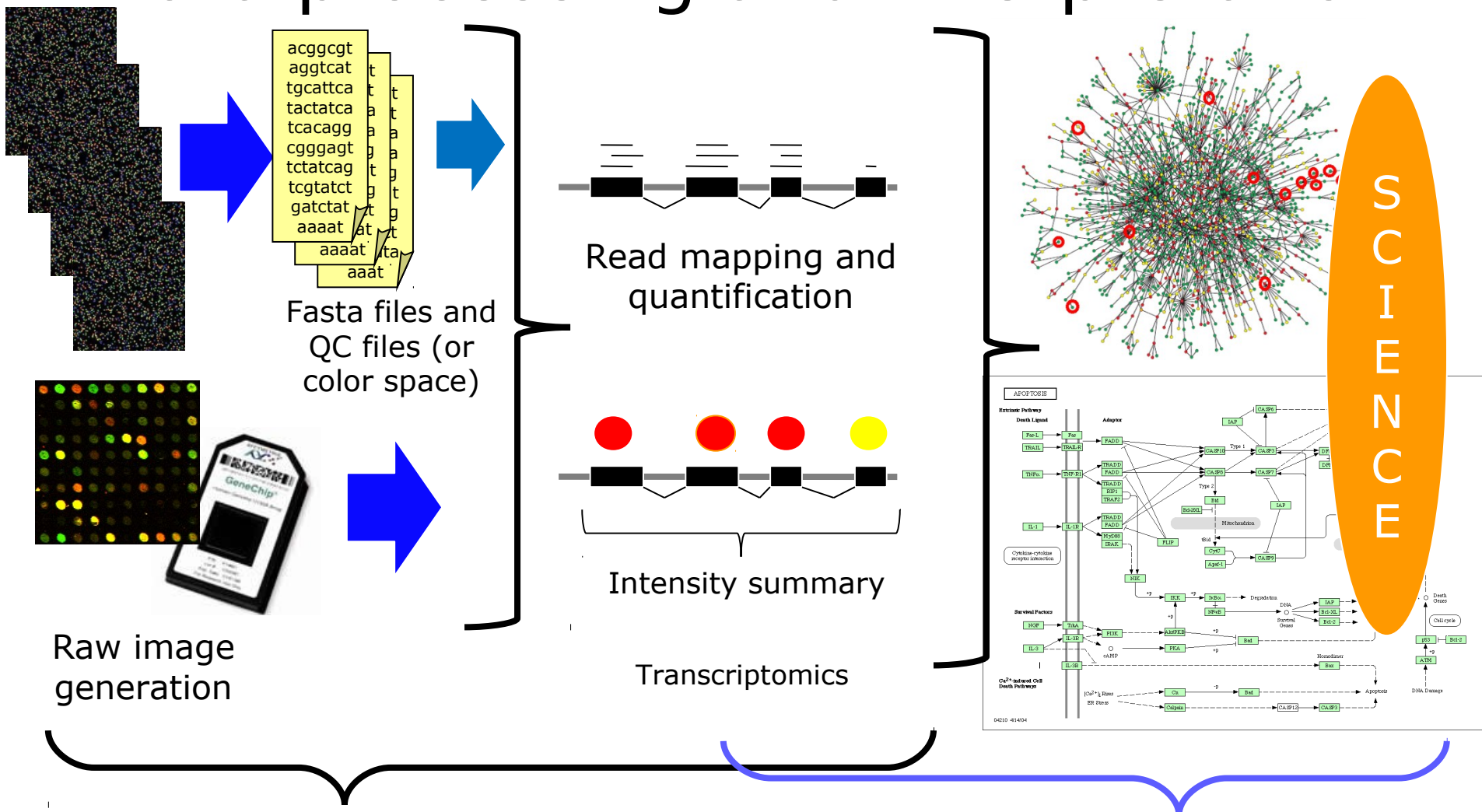


**Source Pubmed. Query:** "high-throughput sequencing"[Title/Abstract] OR "next generation sequencing"[Title/Abstract] OR "rna seq"[Title/Abstract]) AND year[Publication Date]

**Projections 2011** based on January and February



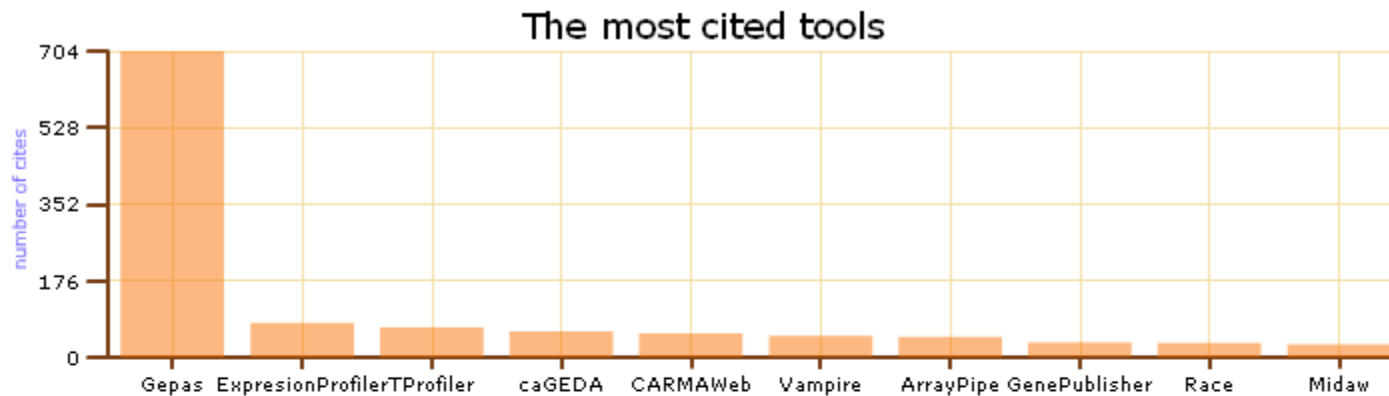
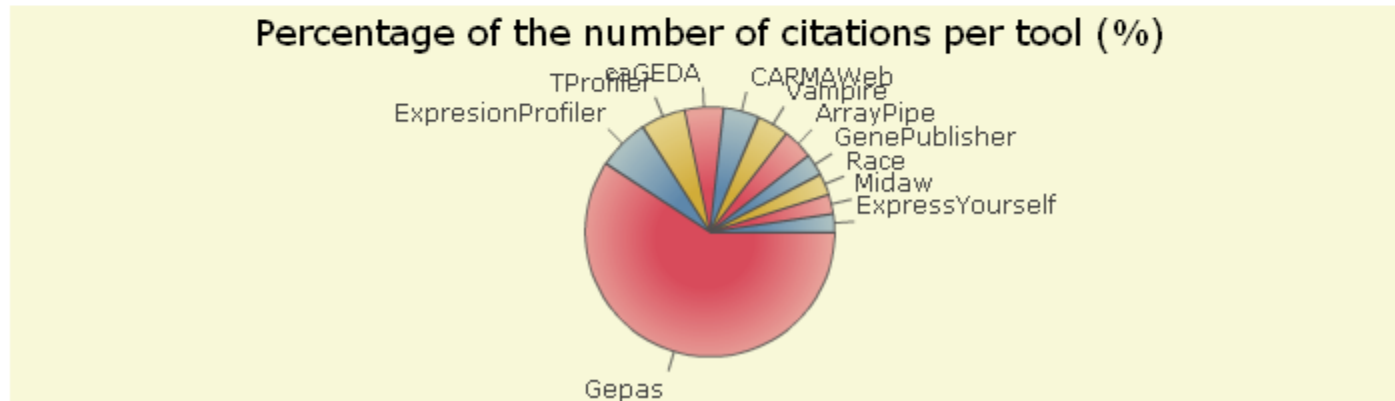
# Genomic data, the double challenge: Data processing and interpretation



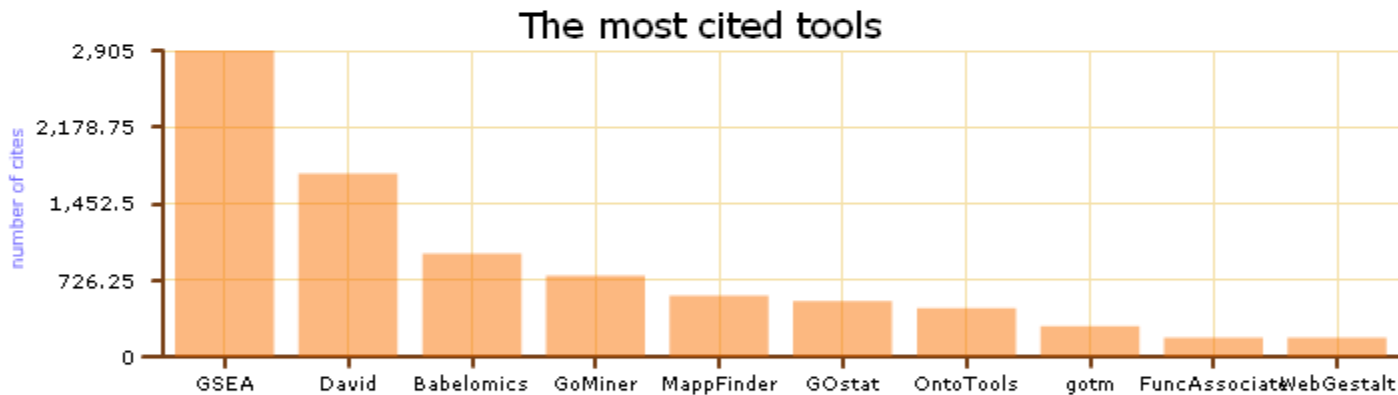
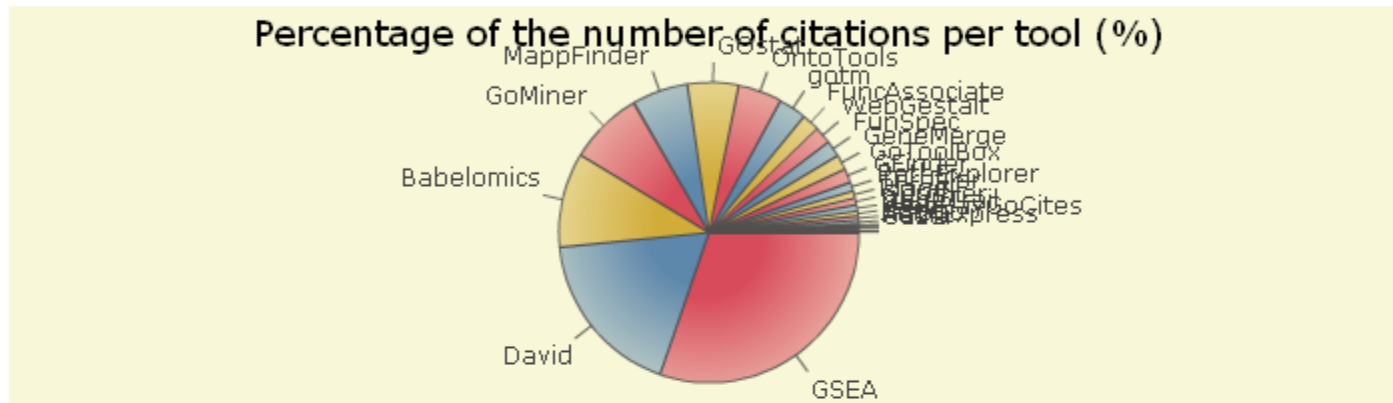
Technology driven

Hypothesis driven

# Tools for gene expression analysis



# Tools for functional profiling



# Some numbers

451 papers cite GEPAS (215 are SOTA cites)

632 papers cite Babelomics (442 are FatiGO cites)

*(source ISI Web of Knowledge, May 2010)*

More than 150,000 experiments analysed during the last year.

More than 1000 experiments per day.



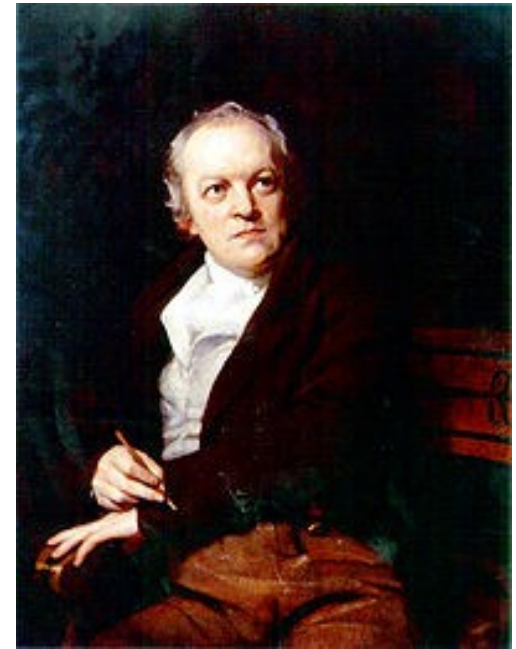




# Background

**The road of excess leads to  
the palace of wisdom**

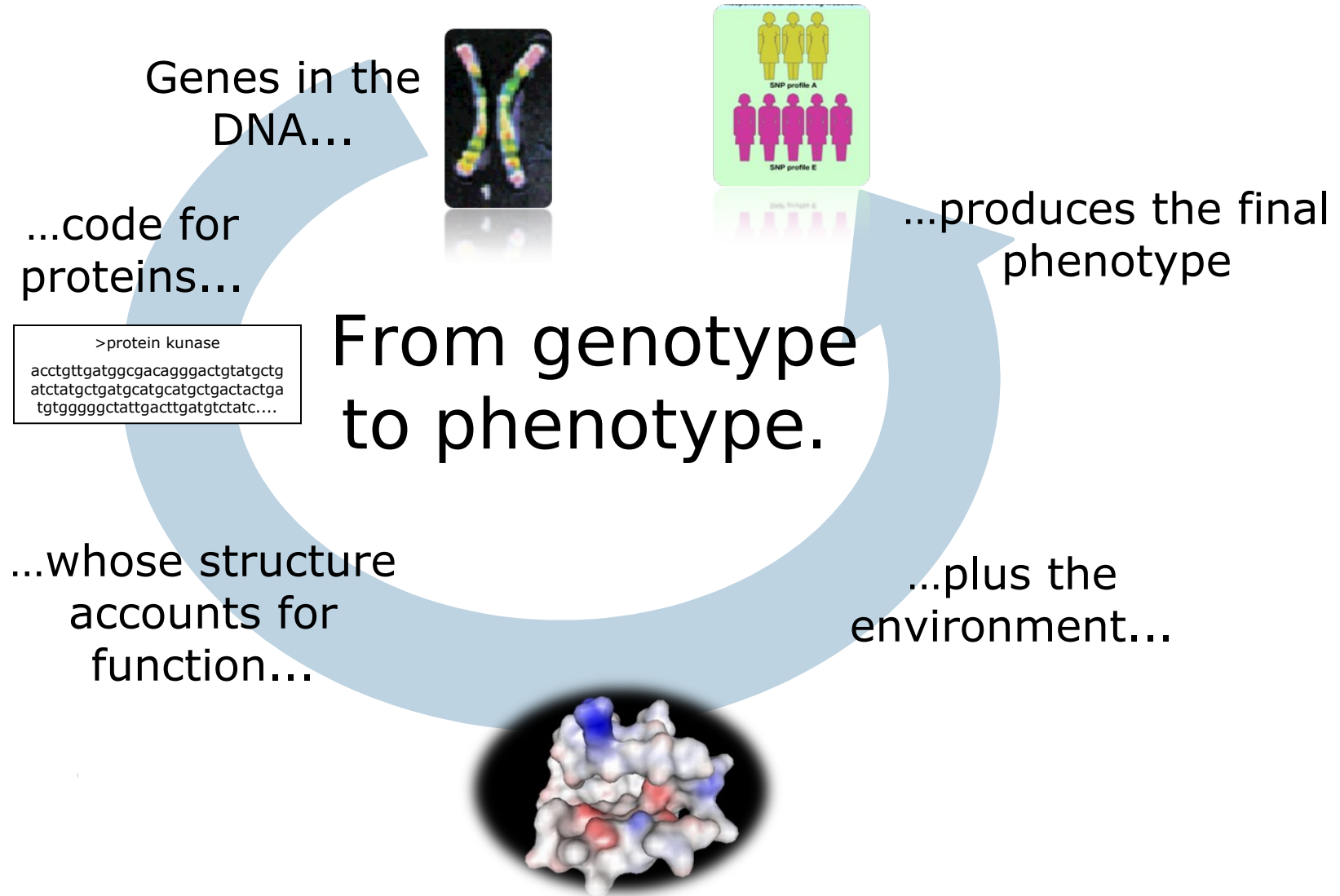
*(William Blake, 28 November 1757 – 12  
August 1827, poet, painter, and printmaker)*



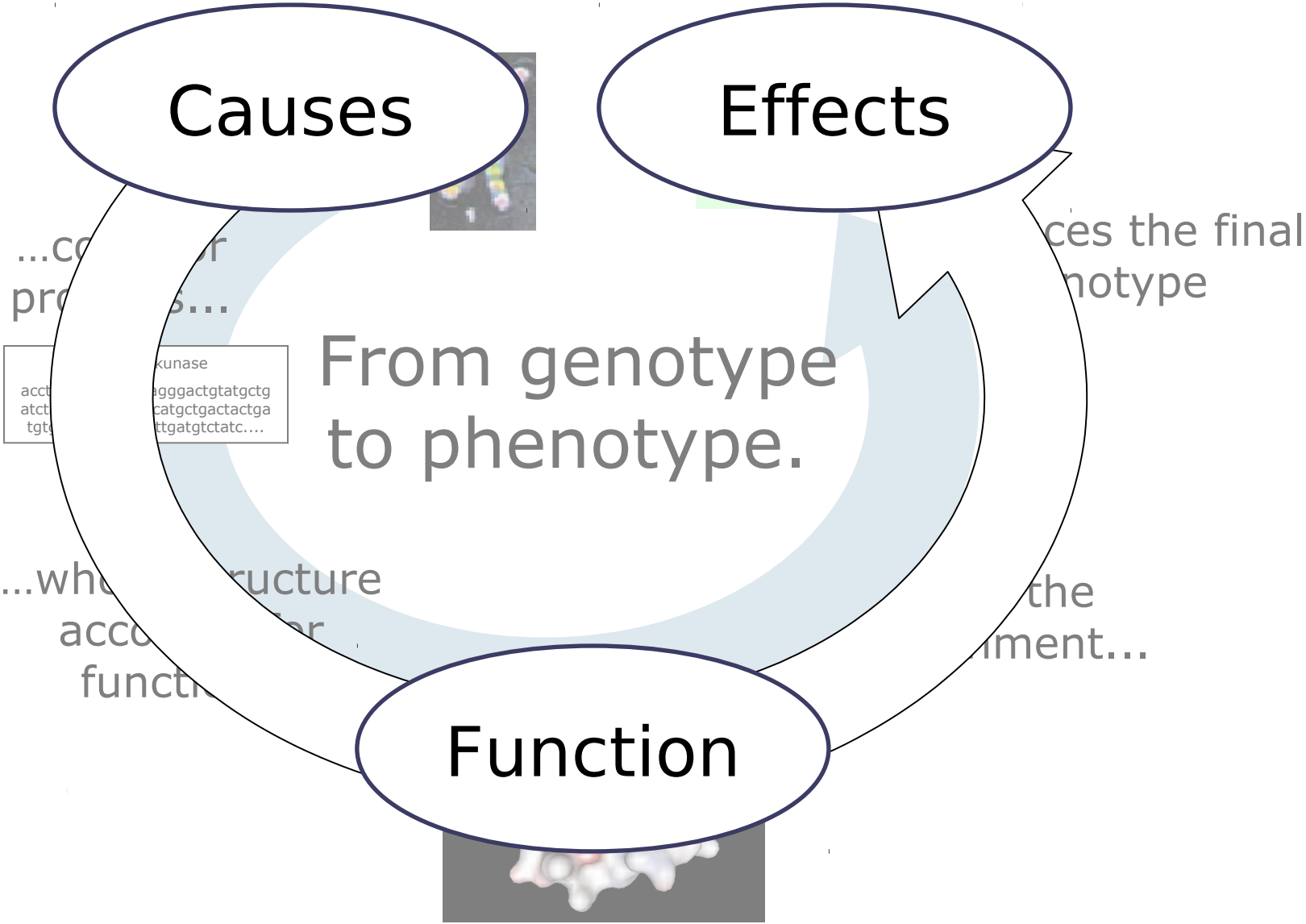
The introduction and popularisation of high-throughput techniques has drastically changed the way in which biological problems **can** be addressed and hypotheses **can** be tested.

But not necessarily the way in which we really address or test them...

# Where do we come from? The pre-genomics paradigm



# Reduccionistic approach to link causes (genome) to effects (phenotype) through actions (function)



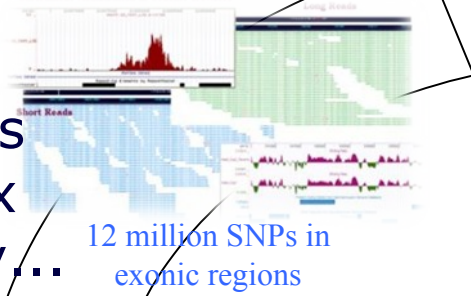


Next Generation Sequencing  
10<sup>9</sup>bp per round

Genes in  
the DNA...

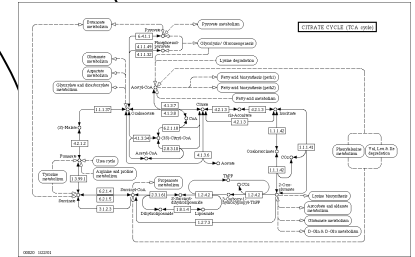


...whose final  
effect  
configures  
the  
phenotype...



# From genotype to phenotype.

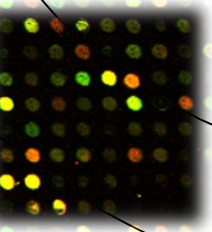
(in the post-genomics scenario)



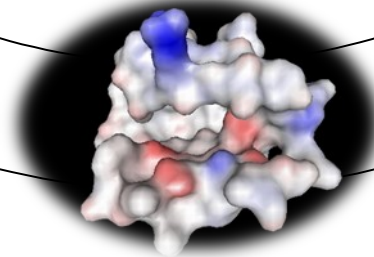
...conforming complex  
interaction networks...

...with its  
complex  
variability...

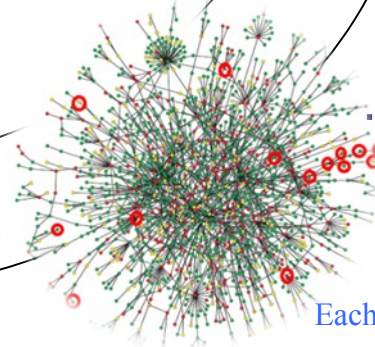
...when they  
are expressed  
in the proper  
moment and  
place...



...code for  
proteins...



That undergo post-translational  
modifications, somatic  
recombination...  
100K-500K proteins



...in cooperation  
with other  
proteins...

Each protein has an average  
of 8 interactions

...that account for  
function if...

Holistic approach. Causes and effects remain essentially the same. The concept of function has changed

Causes

Effects

Function  
(modules of proteins)

From genotype to phenotype

All science is either physics or stamp collecting

Ernst Rutherford

...whose final effect configures the phenotype...

...with its complex variability

Half a million plants individuals

...when

ar  
ir  
n

(in the functional genomics scene)

plex  
KS...

tion  
er

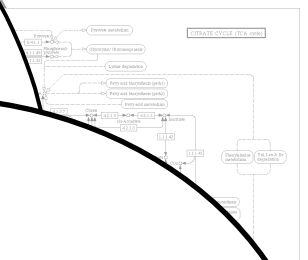
NS...

...code for proteins...

That undergo post-translational modifications, somatic recombination...  
100K-500K proteins

...that account for function if...

...has an average of 8 interactions



# High-throughput data for functional genomics

Genotyping

Genome wide

Metabolomics

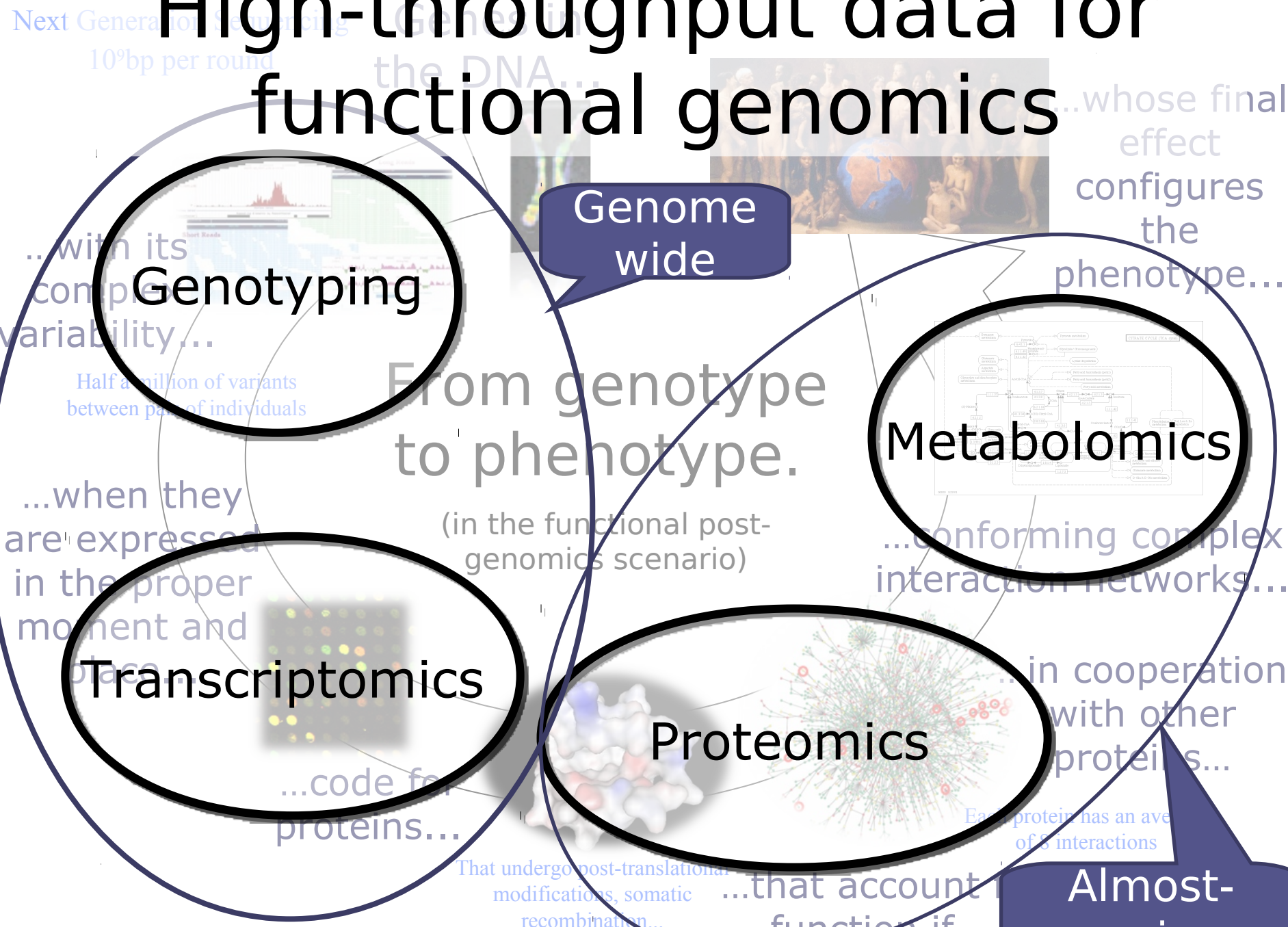
Transcriptomics

Proteomics

Almost-omics

From genotype to phenotype.

(in the functional post-genomics scenario)



# Technologies for transcriptomics and genotyping and the corresponding bioinformatics support



Microarray

User-friendly  
Babelomics

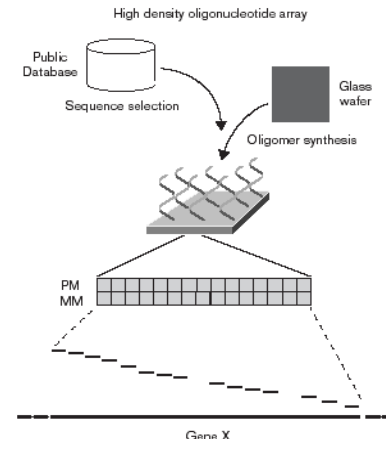
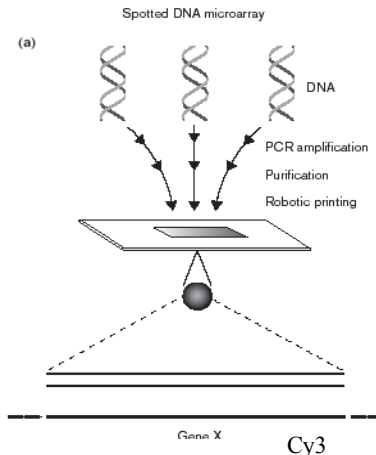
R and  
scripting



NGS

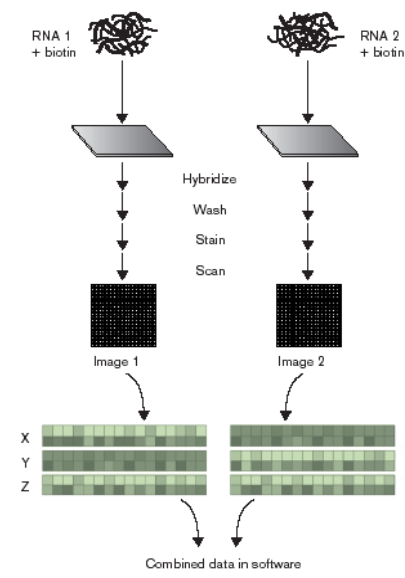
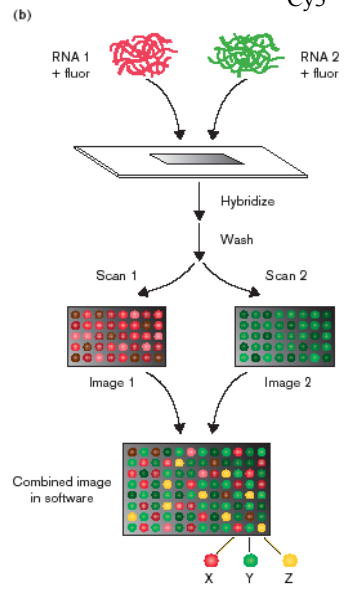


# DNA expression microarrays. Strategies of hybridization



Cy5

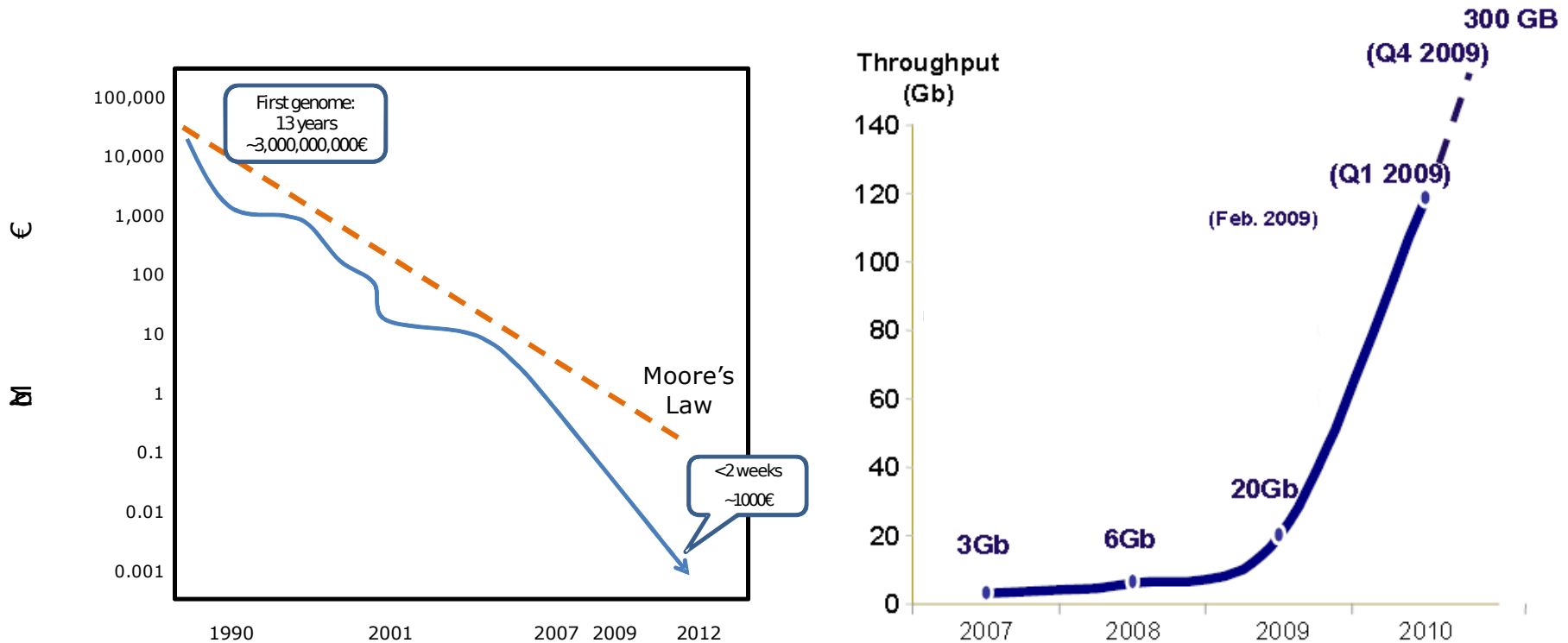
Cy3



Competitive hybridization  
(two colors)

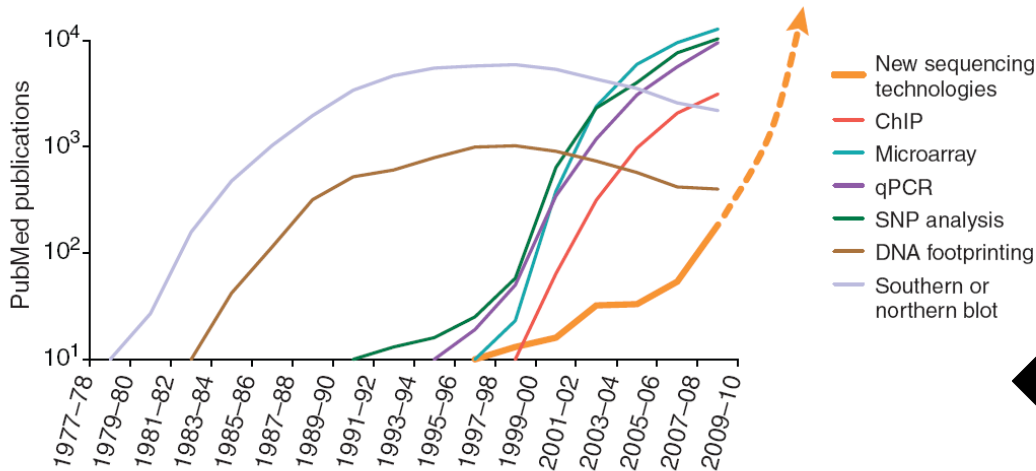
One color

# Next generation sequencing technologies are here

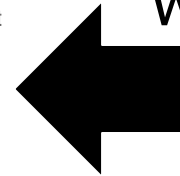


The cost goes down, while the amount of data to manage and its complexity raise exponentially.

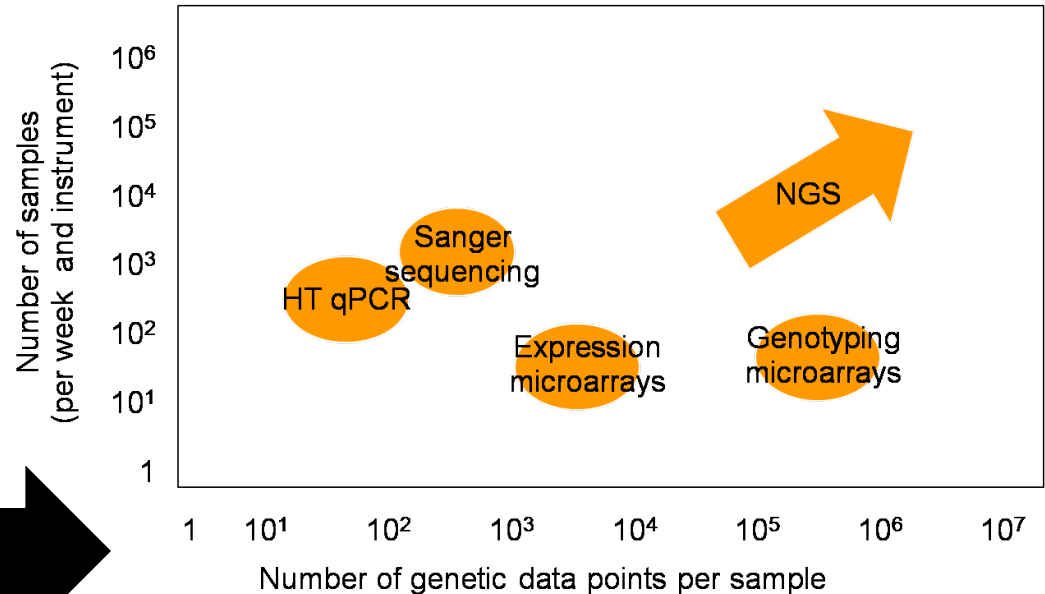
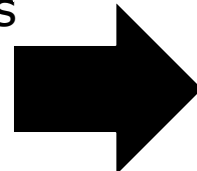
# Next generation sequencing technologies are here



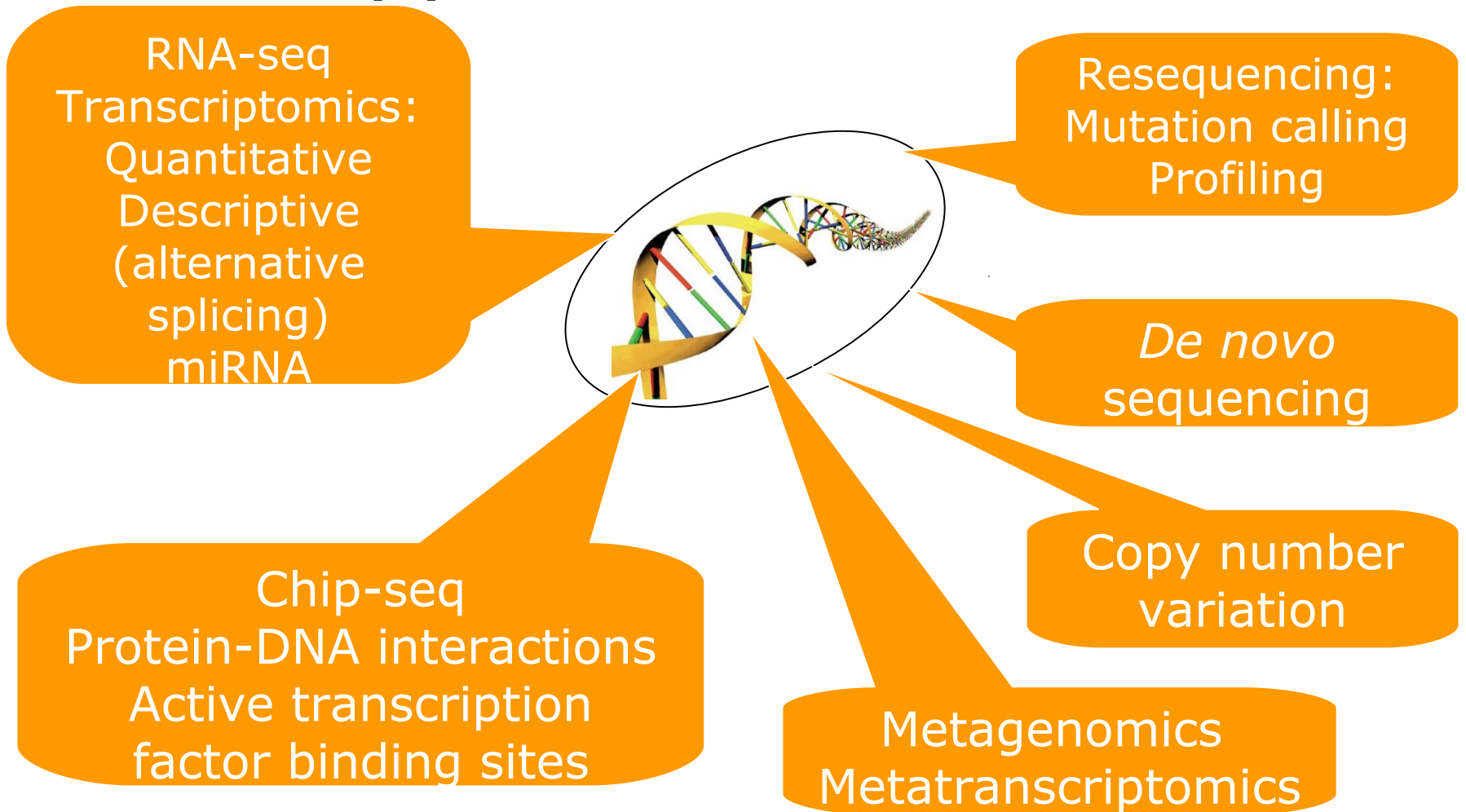
Observed and expected trend of publications in which NGS is being used.



Relative throughput of the different technologies. NGS emerges with a potential of data production that will, eventually wipe out conventional HT technologies in the years coming



# Some of the most common applications of NGS

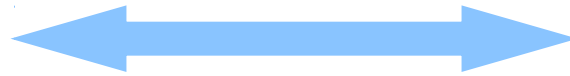
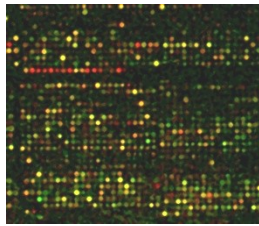




# Gene expression profiling.

## Historic perspective

Differences at phenotype level are the visible cause of differences at molecular level which, in many cases, can be detected by measuring the levels of gene expression. The same holds for different experiments, treatments, strains, etc.



- **Classification of phenotypes / experiments.** Can we distinguish among classes (either known or unknown), values of variables, etc. using molecular gene expression data? (**sensitivity**)
- **Selection of differentially expressed genes** among the phenotypes / experiments. Did we select the relevant genes, all the relevant genes and nothing but the relevant genes? (**specificity**)
- **Biological roles the genes are carrying out in the cell.** What general biological roles are really represented in the set of relevant genes? (**interpretation**)

# Primary analysis

- Transform images corresponding to hybridization intensities (microarrays) or to read counts (NGS) into numbers
- Convert all the measurements to a common scale that makes them comparable across experiments.

# Secondary analysis

Once the measurements are in a common, comparable scale the results can be studied.

Different studies can be made that include class discovery, classification, gene selection, variant calling, etc.

# Studies must be hypothesis driven.

What is our aim? Class discovery? sample classification? gene selection? ...

Can we find groups of experiments with similar gene expression profiles?

Molecular classification of samples

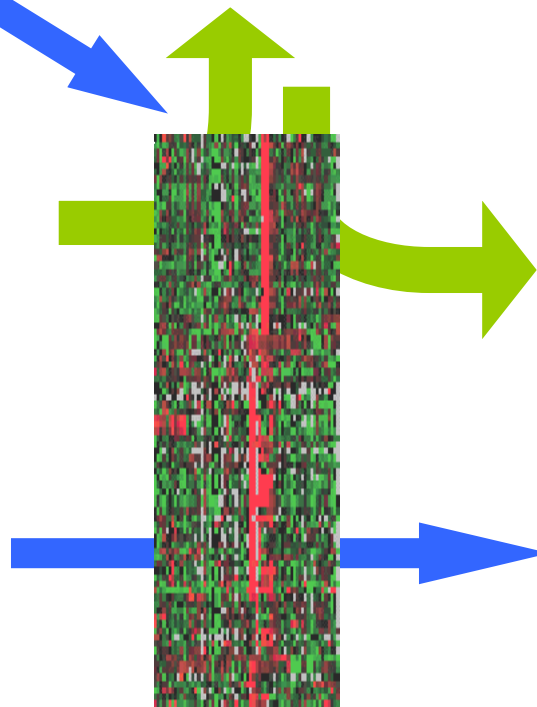
Co-expressing genes...

Different classes...



What genes are responsible for?

What do they have in common?

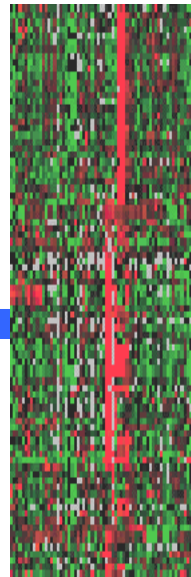




# Unsupervised problem: class discovery

Our interest is in discovering clusters of items (genes or experiments) which we do not know beforehand

Can we find groups of experiments with similar gene expression profiles?



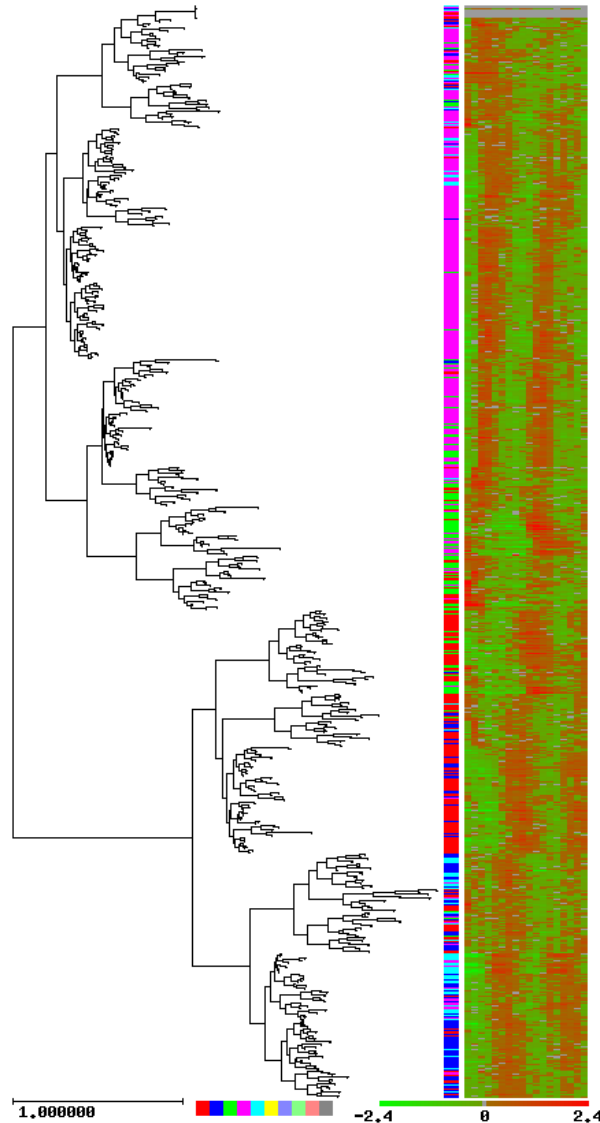
Co-expressing genes...



- What genes co-express?
- How many different expression patterns do we have?
- What do they have in common?
- Etc.



# An unsupervised problem: clustering of genes.



- Gene clusters are previously unknown
- Distance function
- Cluster gene expression patterns based uniquely on their similarities.
- Results are subjected to further interpretation (if possible)



# Clustering of experiments: The problems

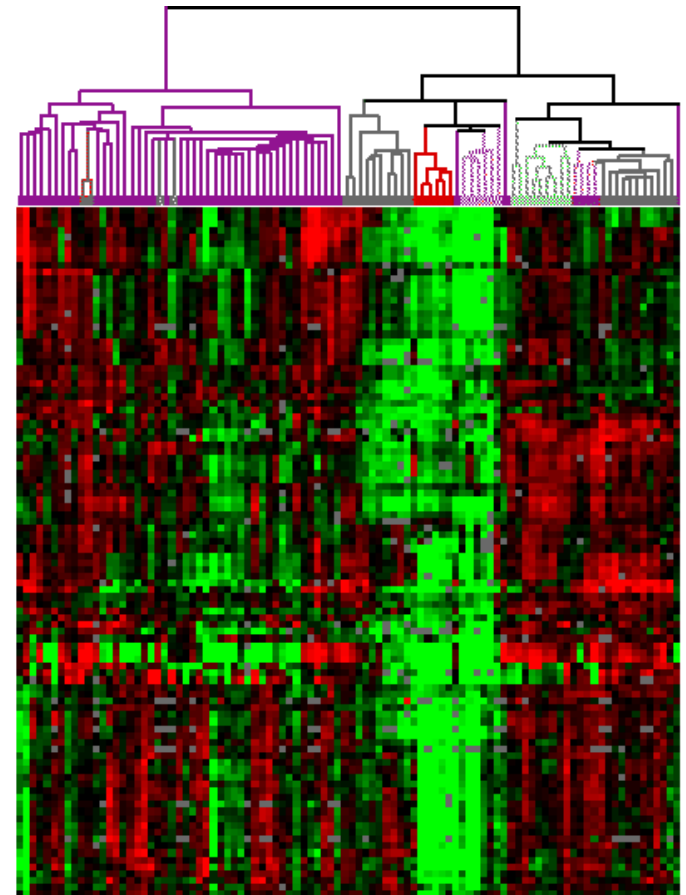
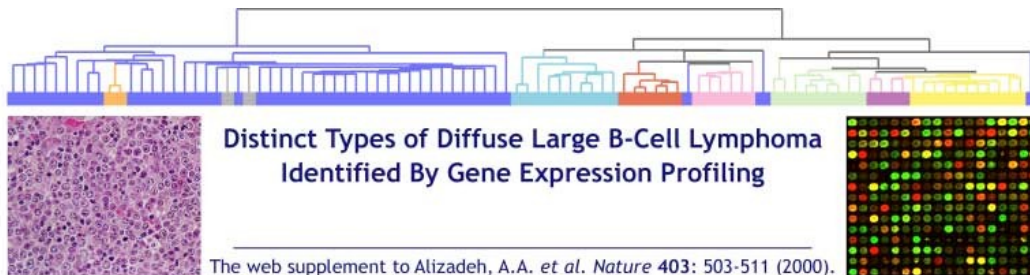
Any gene (regardless its relevance for the classification) has the same weight in the comparison.

If relevant genes are not in overwhelming majority we will find:

Noise

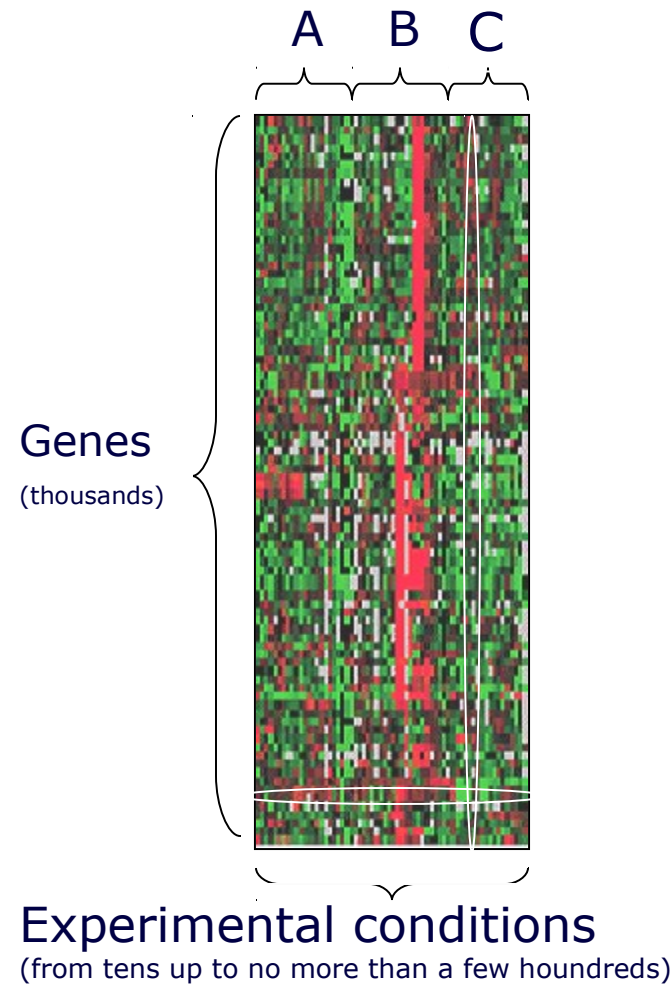
and/or

irrelevant trends



# Supervised problems: Class prediction and gene selection, based on gene expression profiles

Information on classes (defined on criteria external to the gene expression measurements) is used.



Problems:

How can classes A, B, C... be distinguished based on the corresponding profiles of gene expression?

How a continuous phenotypic trait (resistance to drugs, survival, etc.) can be predicted?

And

Which genes among the thousands analysed are relevant for the classification?

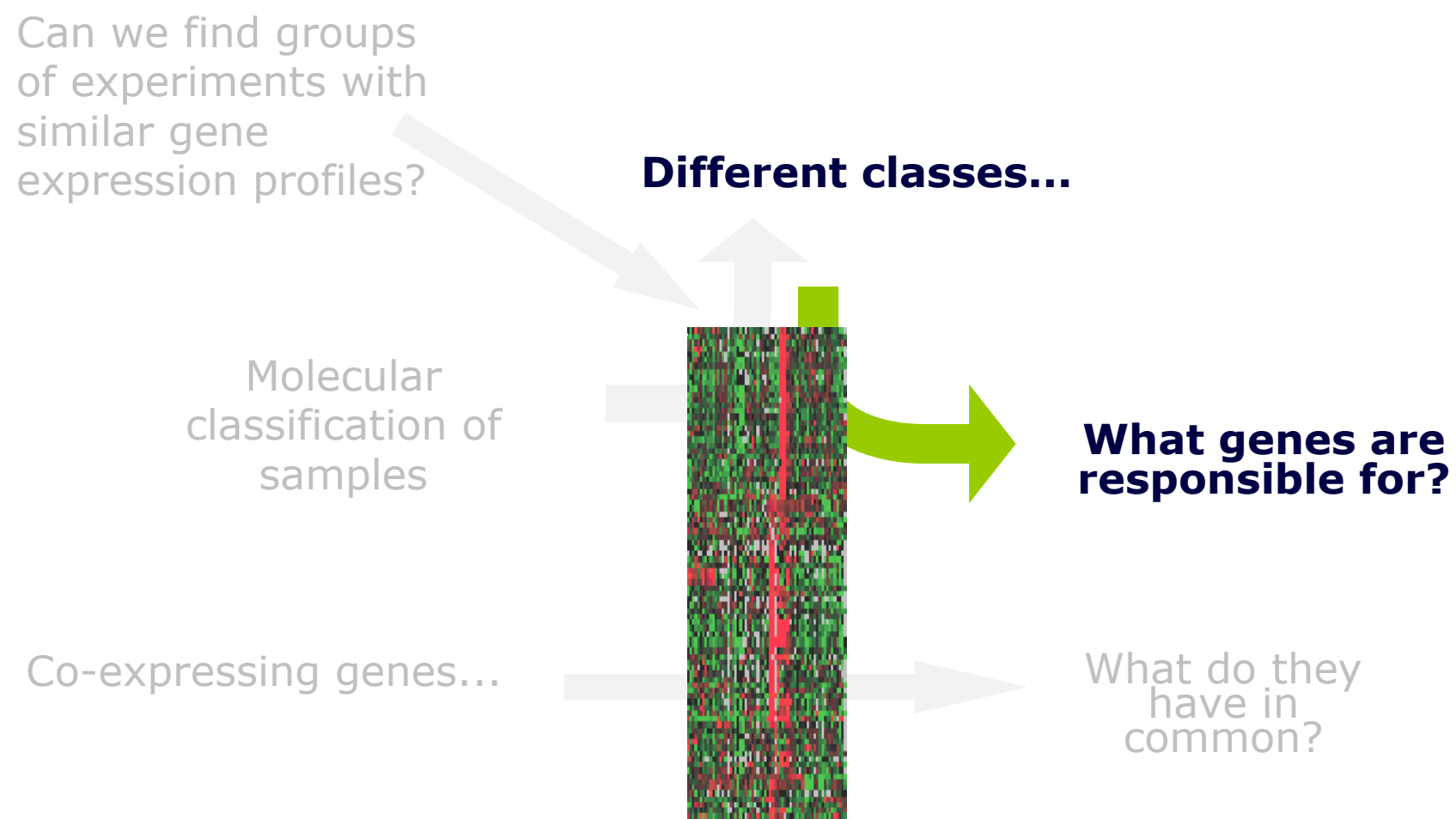
Class prediction

Gene selection



# Studies must be hypothesis driven.

## gene selection



# Gene selection.

The simplest way: univariate gene-by-gene.  
Other multivariate approaches can be used

- **One class**

Limma

- **Two classes**

T-test

Limma

Fold-change

- **Multiclass**

Anova

Limma

- **Continuous variable  
(e.g. level of a  
metabolite)**

Pearson

Spearman

Regression

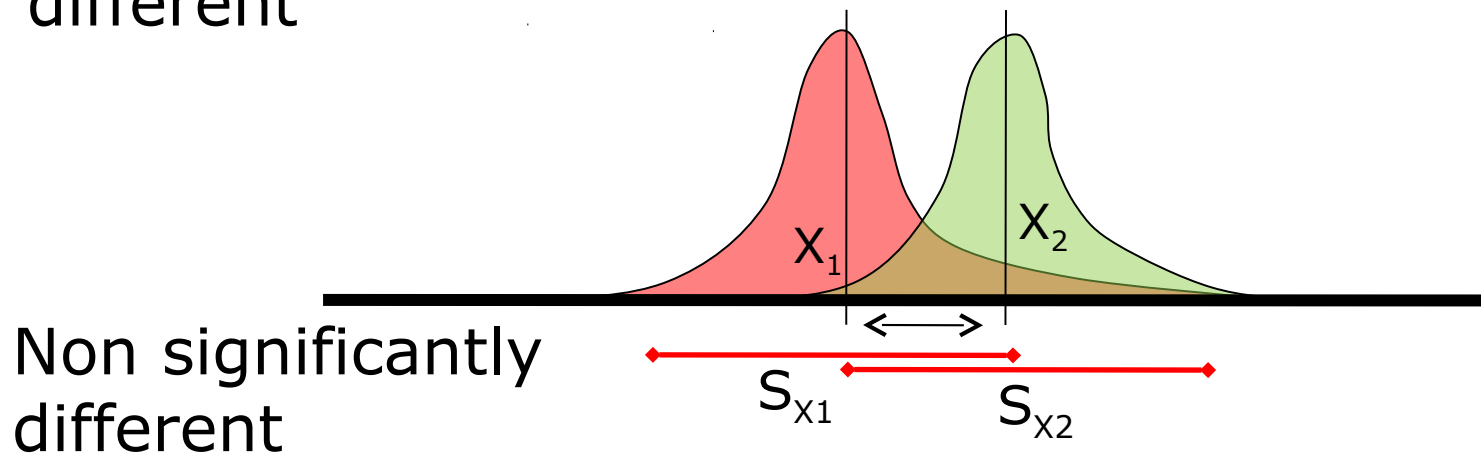
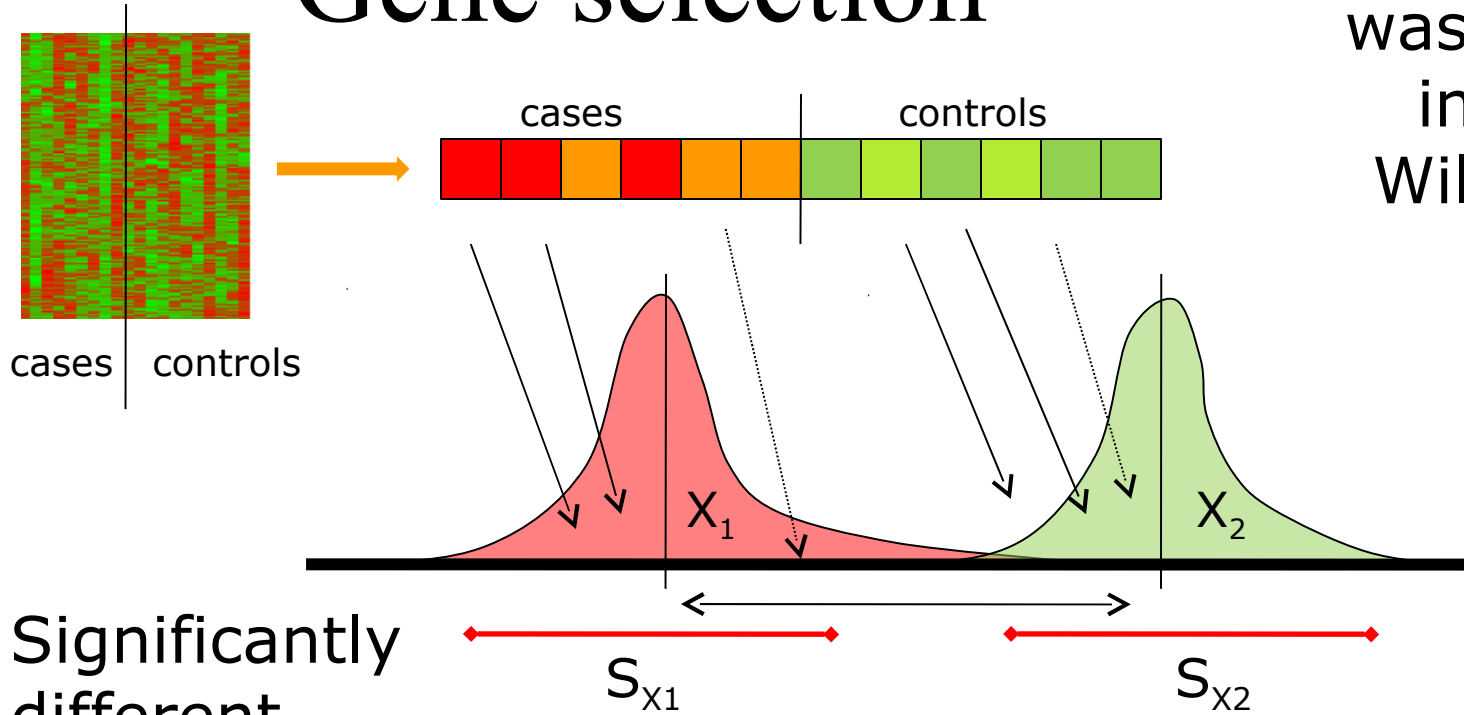
- **Survival**

Cox model

- **Time Course**

# Gene selection

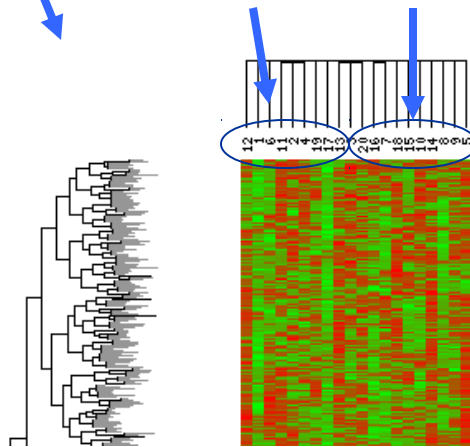
The t-statistic was introduced in 1908 by William Sealy Gosset



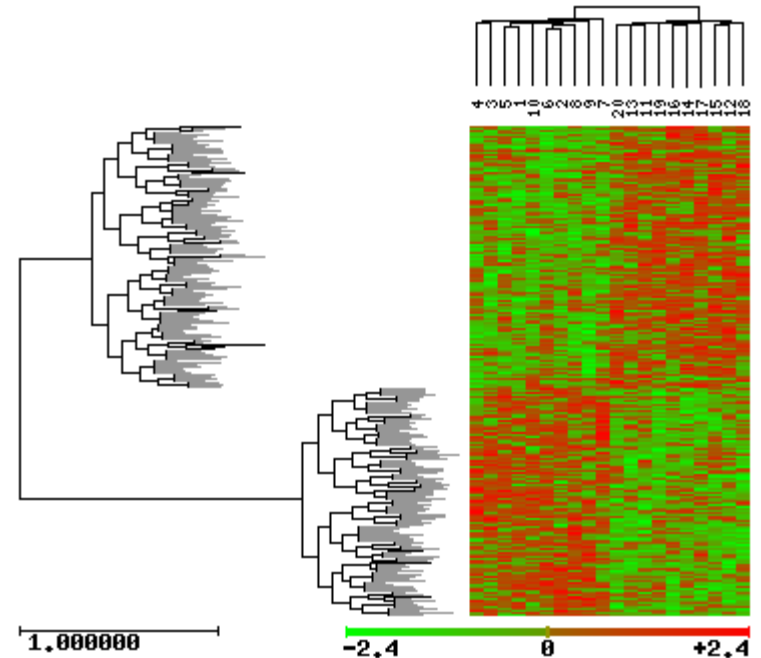
$$t = \frac{\bar{X}_1 - \bar{X}_2}{S_{X_1 X_2} \cdot \sqrt{\frac{2}{n}}} \quad \text{being} \quad S_{X_1 X_2} = \sqrt{\frac{S_{X_1}^2 + S_{X_2}^2}{2}}$$

# A simple problem: gene selection for class discrimination

~15,000 genes  
Case(10)/control(10)



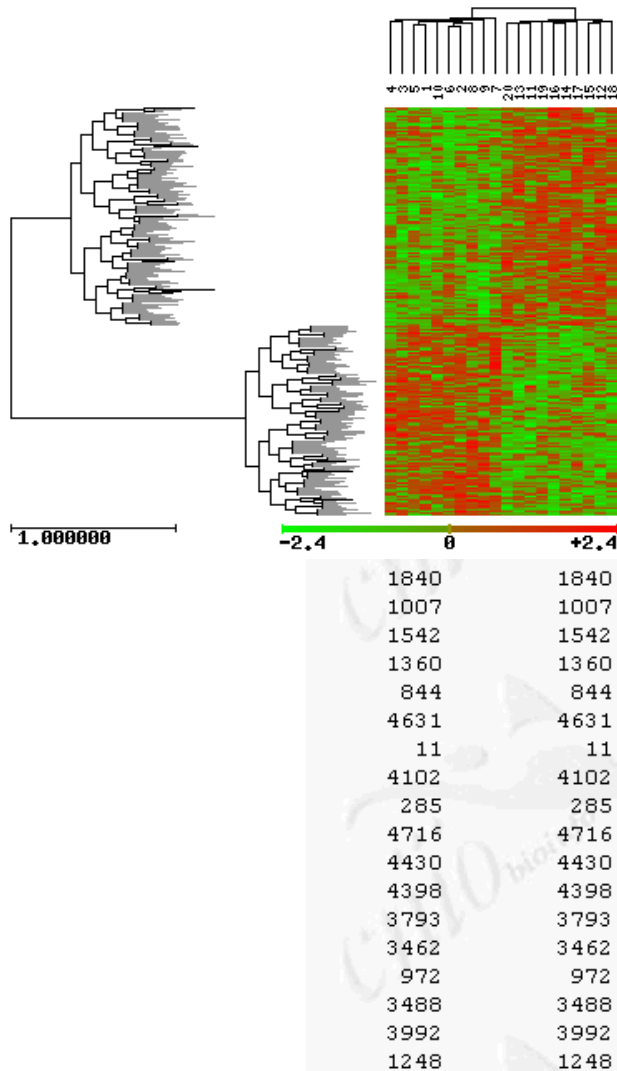
thebest - [04/10/2003 18:57:43 GMT]



Genes differentially expressed  
among classes (t-test), with p-  
value < 0.05

# Sorry... the data was a collection of random numbers labelled for two classes

thebest - [04/10/2003 18:57:43 GMT]



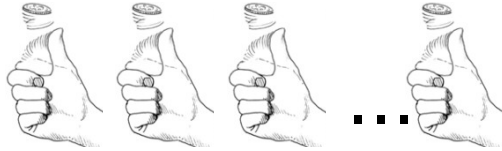
So... Why do we find good p-values?

unadj_p	adj_p	FDR_indep	FDR_dep	obs_stat
0.00019998	0.152685	0.49995	1	5.47044
0.00019998	0.746225	0.49995	1	4.49902
0.0009999	0.983002	0.861025	1	4.01726
0.00149985	0.986401	0.861025	1	3.99374
0.00129987	0.9959	0.861025	1	3.86046
0.00169983	0.9996	0.861025	1	3.7251
0.00169983	0.9996	0.861025	1	3.66628
0.00169983	0.9996	0.861025	1	3.62427
0.00169983	0.9996	0.861025	1	3.60596
0.00169983	0.9996	0.861025	1	3.58109
0.00169983	0.9996	0.861025	1	3.52935
0.00169983	0.9996	0.861025	1	3.43721
0.00169983	0.9996	0.861025	1	3.41937
0.00169983	0.9996	0.861025	1	3.41428
0.00169983	0.9996	0.861025	1	3.4025
0.00169983	0.9996	0.861025	1	3.40212
0.00169983	0.9996	0.861025	1	3.37412
0.00539946	1	0.8888	1	3.36813
0.00219978	1	0.861025	1	3.35909
0.0029997	1	0.861025	1	3.35235
0.00439956	1	0.8888	1	3.28286
0.00669933	1	0.8888	1	3.2427
0.00559944	1	0.8888	1	3.23225
0.00279972	1	0.861025	1	3.22175
0.00429957	1	0.8888	1	3.19595
0.0039996	1	0.8888	1	3.19547
0.0069993	1	0.8888	1	3.12957
0.00849915	1	0.8888	1	3.0987
0.00779922	1	0.8888	1	3.09834

You were not interested *a priori* in the first (whatever), best discriminant, gene.

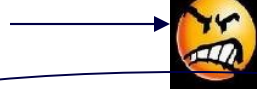
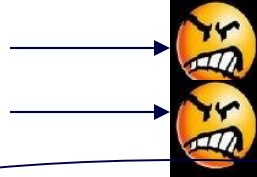
Adjusted p-values must be used!

# On the problem of multiple testing



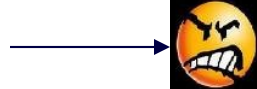
= 10 heads.  $P=0.5^{10}=0.00098$

Take one coin, flip it 10 times. Got 10 heads? Use it for betting



10 heads !!!

⋮



1000 coins

$$P = 1 - (1 - 0.5^{10})^{1000} = 0.62$$

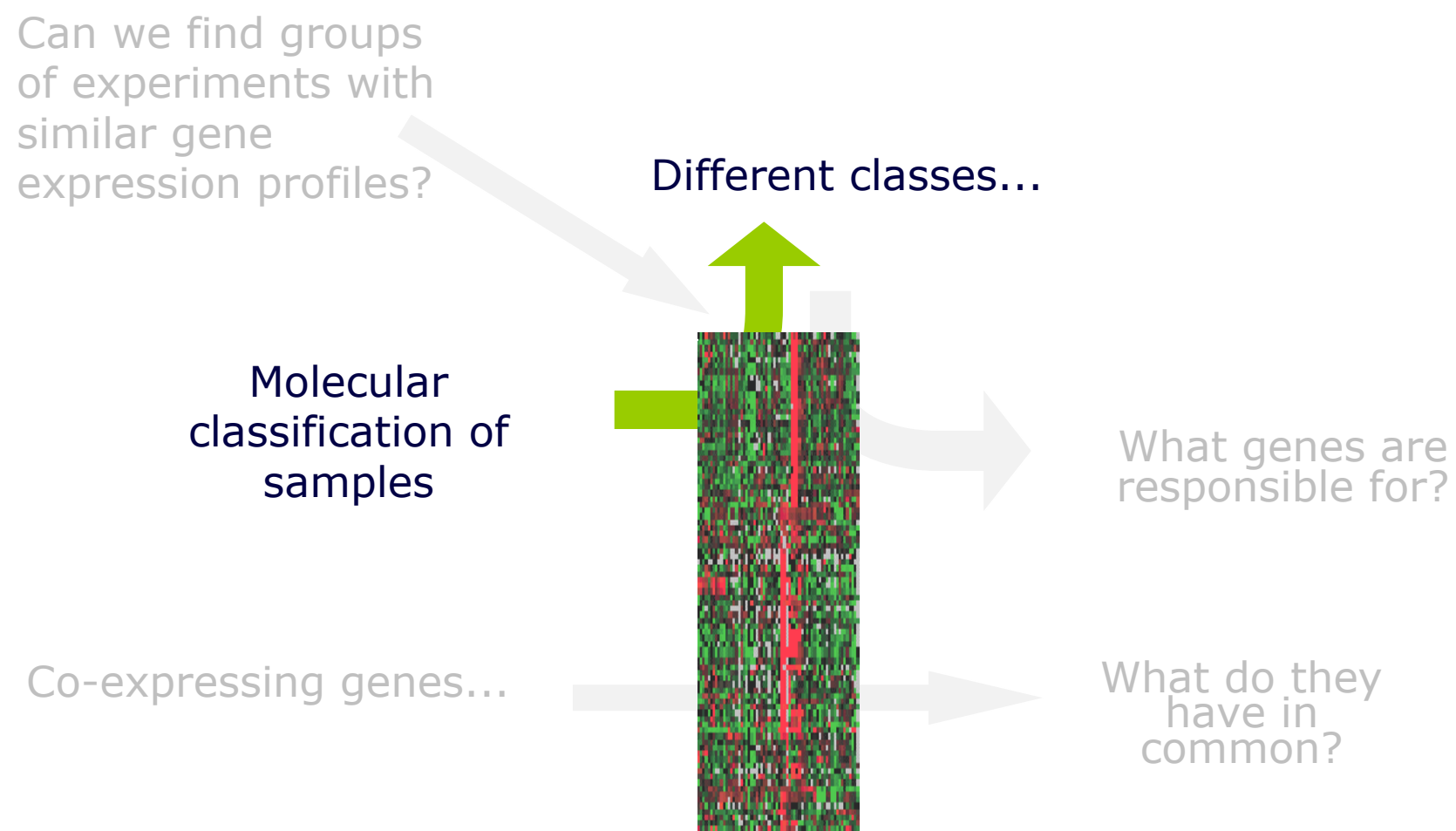
It is not the same getting 10 heads with **my** coin than getting 10 heads in **one among** 1000 coins

Will you still use this coin for betting?



# Studies must be hypothesis driven.

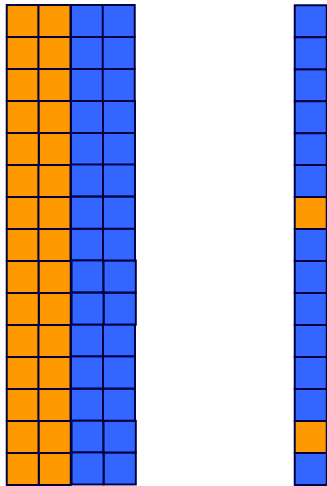
## sample classification



# Of predictors and molecular signatures

What is a predictor?

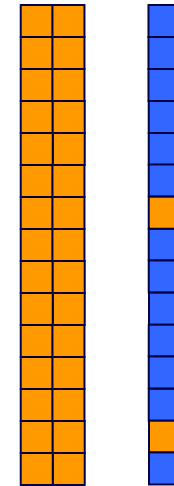
A B X



Is X, A  
or B?

$$\text{Diff (B, X)} = 2$$

Intuitive notion:



$$\text{Diff (A, X)} = 13$$

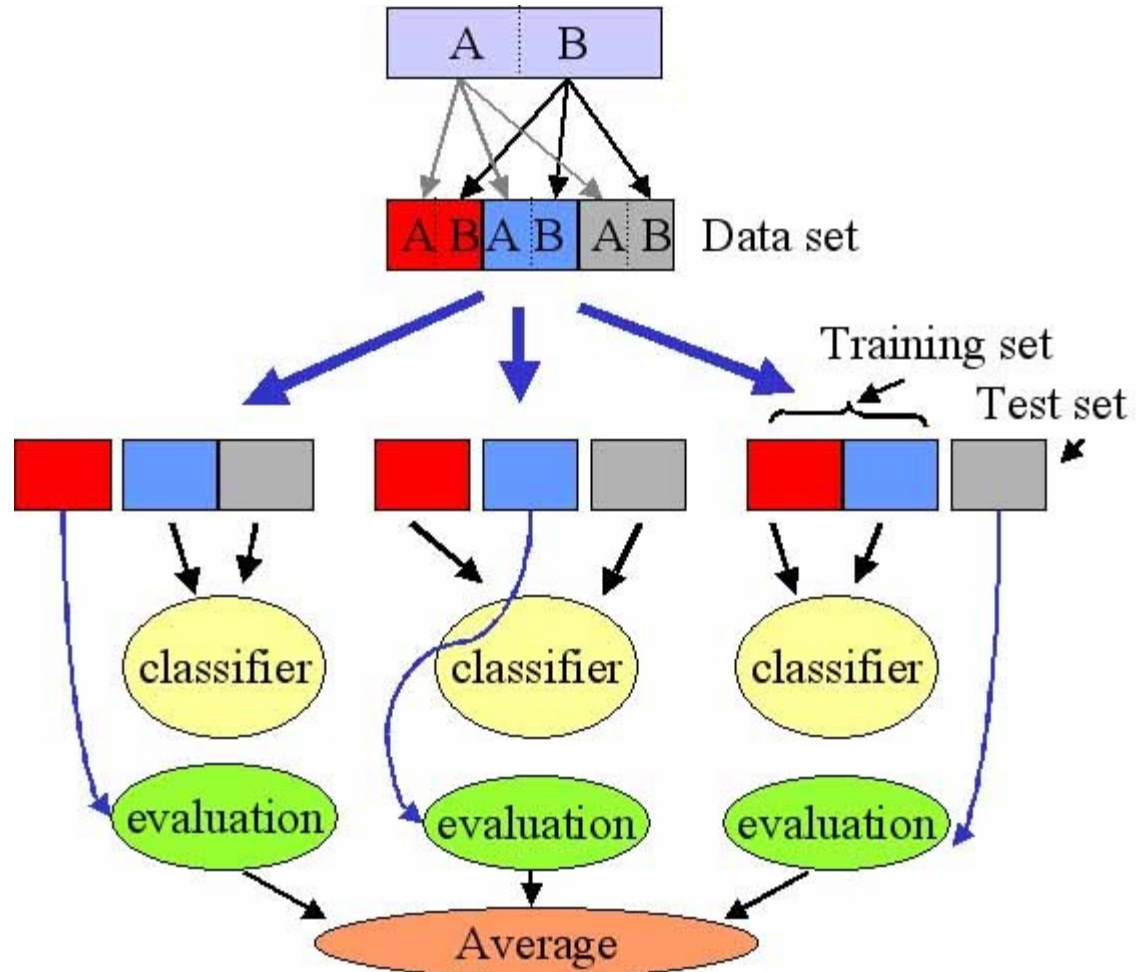
Most probably X belongs to class B

Algorithms: DLDA, KNN, SVM, random forests, PAM, etc.

# Cross-validation

The efficiency of a classifier can be estimated through a process of cross-validation.

Typical are three-fold, ten-fold and leave-one-out (LOO), in case of few samples for the training



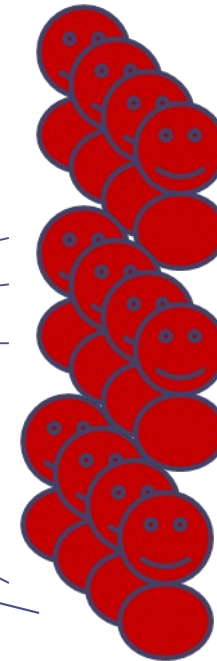
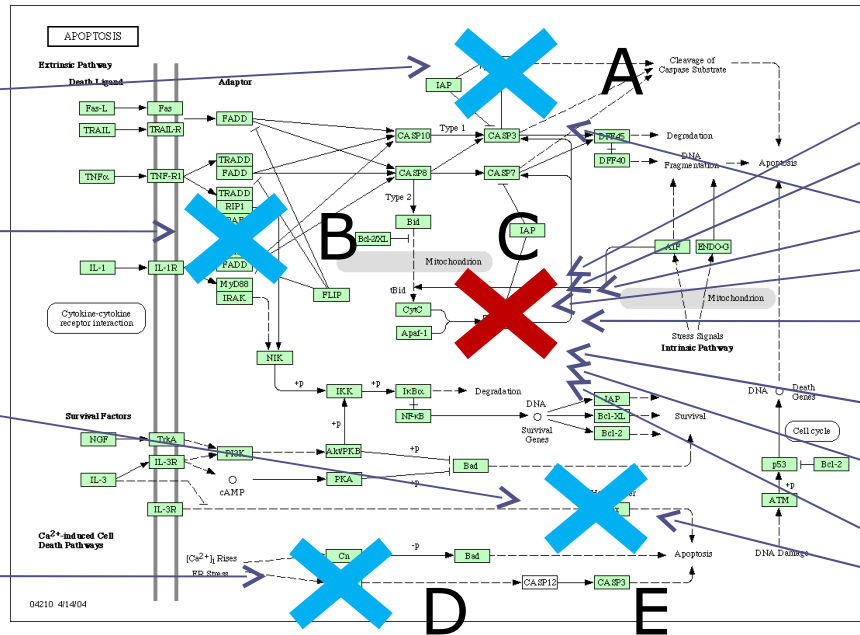
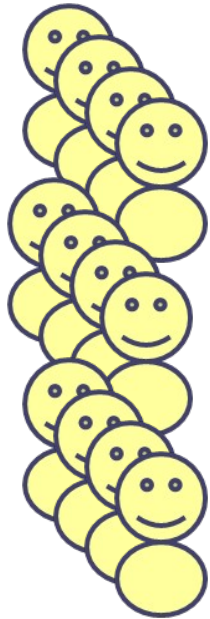






# Genotyping to find mutations associated to diseases

The simplest case: monogenic disease

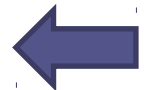


Controls

Cases

Gene A	1	0	0	0	0	0	0	0	0	0	0	0	0	0
Gene B	0	0	0	1	0	0	0	0	0	0	0	0	0	0
Gene C	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Gene D	0	0	0	0	0	0	0	0	0	0	0	0	1	0
Gene E	0	0	0	0	0	1	0	0	0	0	0	0	0	0

	0	0	0	0	1	0	0	0	0	0	0	0	0	0
	0	0	0	0	0	0	0	0	0	0	0	0	0	0
	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	0	0	0	0	0	0	0	0	0	0	0	0	0	0
	0	0	0	0	0	0	0	0	1	0	0	0	0	0



# The real life in GWAS

Our analysis of Hirschsprung's disease

54 trios of short-segment Hirschsprung's disease Affy 6.0 (1million SNPs)

Conventional TDT test reports only 4 significant SNPs mapping only on one gene: RET, already knowk to be associated to the disease

This is not a matter of sample size: an example of GWAS in Breast Cancer.

The CGEMS initiative. (Hunter et al. Nat Genet 2007)

1145 cases 1142 controls. Affy 500K

Conventional association test reports only significant 4 SNPs mapping only on one gene: FGFR2

Conclusions: conventional tests are not providing much resolution.

What is the problem with them? Are there solutions?





# Functional profiling of genome-scale experiments in the post-genomic era

My data...

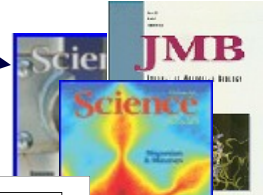
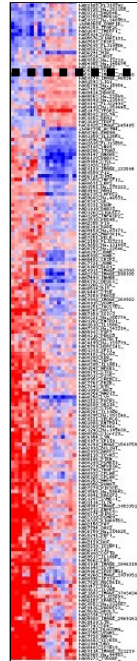
How are structured?

What are these groups?

What is this gen?

	E	F	G	H	I	J	K	L	M	N
65	578.6*		1.4	0.26	M12481	Mouse cytoplasmic beta-actin mRNA (5, M, 3)	represe			
66	534.9*	~1.6		0.22	M12481	Mouse cytoplasmic beta-actin mRNA (5, M, 3)	represe			
67	403.6*	~1.5		0.15	X61366	SGD: YELU02C	Yeast S. cerevisiae WBP1 Oligosaccharyl			
68	535.2*	~1.6		0.22	U18530	SGD: YELU16W	Yeast S. cerevisiae Protein of unknown fu			
69	-567.7*	~1.6		-0.27	M23316	SGD: YELU24C	Yeast S. cerevisiae RIP1 Rieske non-sulfur			
70	-114.5*	~1.1		-0.03	K02207	SGD: YELU21W	Yeast S. cerevisiae URA3 gene coding for			
71	-125.4*		-1	-0.01	Cluster Incl M16465	Calpactin I light chain /cds=(89,361) /gb=M16465				
72	1091.6		-1.2	-0.14	Cluster Incl Z87446	M. musculus spermidine synthase gene /cds=(1,12)				
73	-757.0		-1.3	-0.17	Cluster Incl X12973	Mouse MLC1/FMLL3P gene for myosin alkali				
74	9636.6		1.3	0.63	Cluster Incl AB49035	U1-M-BH1-aga-a06-0-U1 s1	Mus musculus c			
75	-847.4		-1.3	-0.21	Cluster Incl AW123542	U1-M-BH2-1-aga-f01-0-U1 s1	Mus musculus c			
76	2563.1		1.1	0.09	Cluster Incl AF055983	Mus musculus proteasome alpha7/08 subu				
77	192.5*	~1.2		0.05	Cluster Incl AB006361	Mus musculus mRNA for prostaglandin D s				
78	2990.2*	~1.4		1.63	Cluster Incl AB006361	Mus musculus mRNA for prostaglandin D s				
79	-20.1		-1		Cluster Incl AB011081	Mus musculus mRNA for huntingtin intera				
80	1380.9*	~2.6		1.81	Cluster Incl AB011081	Mus musculus mRNA for huntingtin intera				
81	753.2*		1.2	0.1	Cluster Incl U97170	Mus musculus protein kinase inhibitor gamma				
82	-2774.7*	~1.9		-1.43	Cluster Incl M36120	Keratin complex 1, acidic, gene 19 /cds=(0,12)				
83	3614.4*	~5.1		1.98	Cluster Incl U19604	DNA ligase 1, ATP-dependent /cds=(304,3054)				
84	0*	~0.0		0	Cluster Incl AB51492	U1-M-BH3-aga-d04-0-U1 s2	Mus musculus c			
85	3310.9		1.2	0.24	Cluster Incl AB025408	Mus musculus mRNA for sid470p, complet				
86	-1291	~1.5		-0.42	Cluster Incl AF059735	Mus musculus C-terminal binding protein 2				
87	-263.3*	~1.3		-0.09	Cluster Incl AF053454	Mus musculus tetraspan TM4SF (Tspan-6)				
88	77.5*		1.1	0.01	Cluster Incl D45850	Hydroxysteroid 17-beta dehydrogenase 1 /cds				
89	2047.2*	~3.3		1.1	Cluster Incl AF039299	Mus musculus 17-beta-hydroxysteroid de				
90	809.9*	~1.9		0.38	Cluster Incl M04487	Vascular cell adhesion molecule 1 /cds=(57, 62)				
91	-124.3*	~1.1		-0.03	Cluster Incl U12884	Mus musculus C57BL/6 vascular cell adhesio				
92	-675.5*	~1.8		-0.37	Cluster Incl U12884	Mus musculus C57BL/6 vascular cell adhesio				
93	1465.4*	~2.7		0.76	Cluster Incl A123836	Mus musculus mRNA for nucleoside diphos				
94	838.2		1.1	0.1	Cluster Incl U70475	Nuclear, factor, erythroid derived 2, like 2 /cds				
95	4939.4*	~6.7		8.84	Cluster Incl AF045673	Mus musculus F1J-LRR associated protein-				
96	148.3*	~1.2		0.04	Cluster Incl AB91475	U1-M-BH3-aga-x1	Mus musculus cDNA, 3' end /cd			

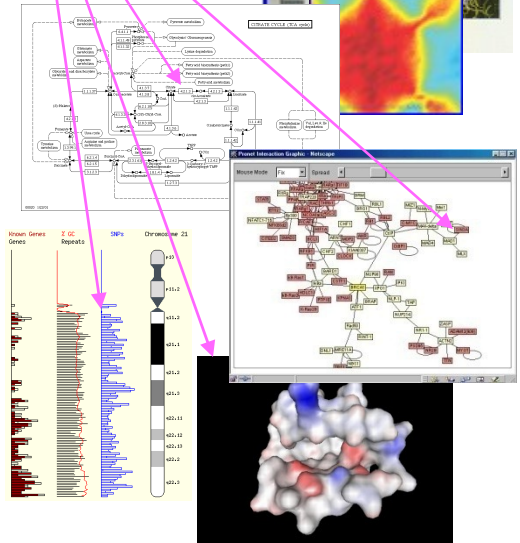
A B



Cell cycle...



- I19380: Calmodulin 3 /cds=(109,558) /gb=M19380 /gi=469419
- AB42320: U1-M-AH1-aga-b-11-0-U1 s1
- A1242683: Mus musculus mRNA for calpactin 2 precursor (cta
- U126204: Dipeptidylpeptidase 4 /cds=(117,2399) /gb=U12620 /g
- M13444: Mouse alpha-tubulin isoform M-alpha-4 mRNA, compl
- U11027: Mus musculus C57BL/6J Srd61 protein complex gam
- J03926: Phosphofruktokinase, liver, B-type /cds=(42,2394) /gb
- Z87745: M. musculus mRNA for phosphatase 2A catalytic subu
- U80932: Serine/threonine kinase 6 /cds=(48,1235) /gb=U80932
- U47024: Maternal embryonic message 3 /cds=(137,2401) /gb=
- AF075136: Mus musculus SimA-associated protein (sp30) mR
- M25944: Mouse carbonic anhydrase II (CAII) mRNA, 3' end /cd
- X74671: Neurofibromin 2 /cds=(676,2366) /gb=X74671 /gi=
- M12649: Mouse myb proto-oncogene mRNA encoding 71 kd m
- AW125458: U1-M-BH2-2-aga-a07-0-U1 s1
- Mus musculus cDN
- U84903: Ribosomal protein L23 /cds=(61,501) /gb=U84903 /gi=
- U35141: Mus musculus retinoblastoma-binding protein (mRb) g
- U19521: Mus musculus vesicle transport protein (nucl-15) ml
- M15268: Aminolevulinic acid synthase 2, erythroid /cds=(0,179)
- M25149: Tissue specific transplantation antigen P91A /cds=(0,
- X66449: Calcyclin /cds=(159,428) /gb=X66449 /gi=50271 /gi=



Analysis

Functional profiling

Links

- Aedes aegypti**  
home page | site map
- Anopheles gambiae**  
home page | site map
- Bos taurus**  
home page | site map
- Caenorhabditis elegans**  
home page | site map
- Canis familiaris**  
home page | site map
- Cavia porcellus**  
home page | site map
- Ciona intestinalis**  
home page | site map
- Ciona savignyi**  
home page | site map
- Danio rerio**  
home page | site map
- Dasybus novemcinctus**  
home page | site map
- Drosophila melanogaster**  
home page | site map
- Microcebus murinus**  
home page | site map
- Monodelphis domestica**  
home page | site map
- Mus musculus**  
home page | site map
- Myotis lucifugus**  
home page | site map
- Ochotona princeps**  
home page | site map
- Ornithorhynchus anatinus**  
home page | site map
- Oryctolagus cuniculus**  
home page | site map
- Oryzias latipes**  
home page | site map
- Otolemur garnettii**  
home page | site map
- Pan troglodytes**  
home page | site map
- Pongo pygmaeus**  
home page | site map
- Echinops telfairi**  
home page | site map
- Equus caballus**  
home page | site map
- Erinaceus europaeus**  
home page | site map
- Felis catus**  
home page | site map
- Gallus gallus**  
home page | site map
- Gasterosteus aculeatus**  
home page | site map
- Homo sapiens**  
home page | site map
- Loxodonta africana**  
home page | site map
- Macaca mulatta**  
home page | site map

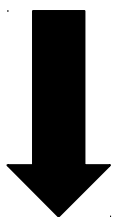
**Genome  
Annotation**

**Structural Annotation    Functional  
Annotation**

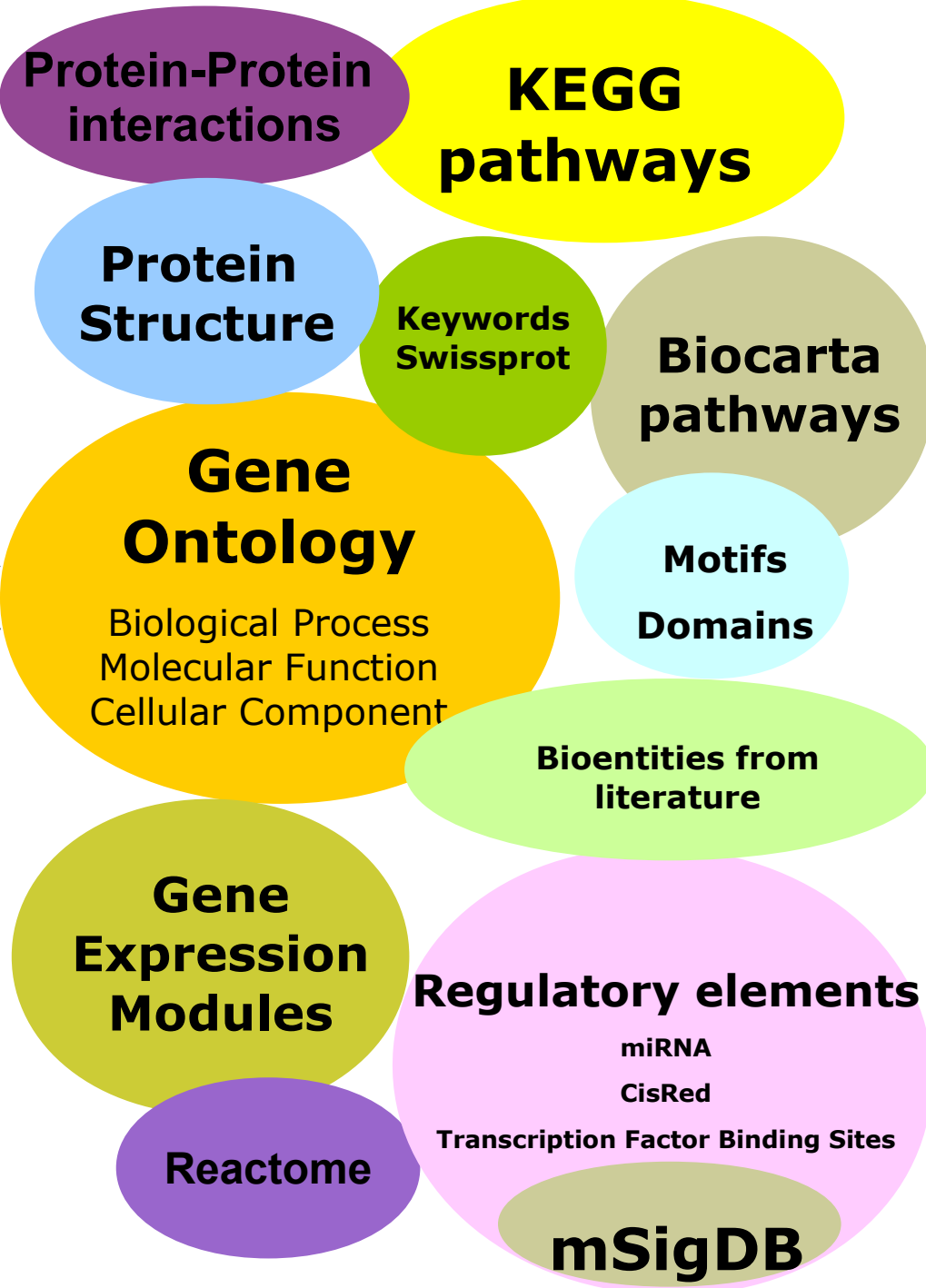
**Biological  
Databases**



**Gene Annotation**



**Gene Set Annotation**



**Protein-Protein  
interactions**

**KEGG  
pathways**

**Protein  
Structure**

**Keywords  
Swissprot**

**Biocarta  
pathways**

**Gene  
Ontology**  
Biological Process  
Molecular Function  
Cellular Component

**Motifs  
Domains**

**Bioentities from  
literature**

**Gene  
Expression  
Modules**

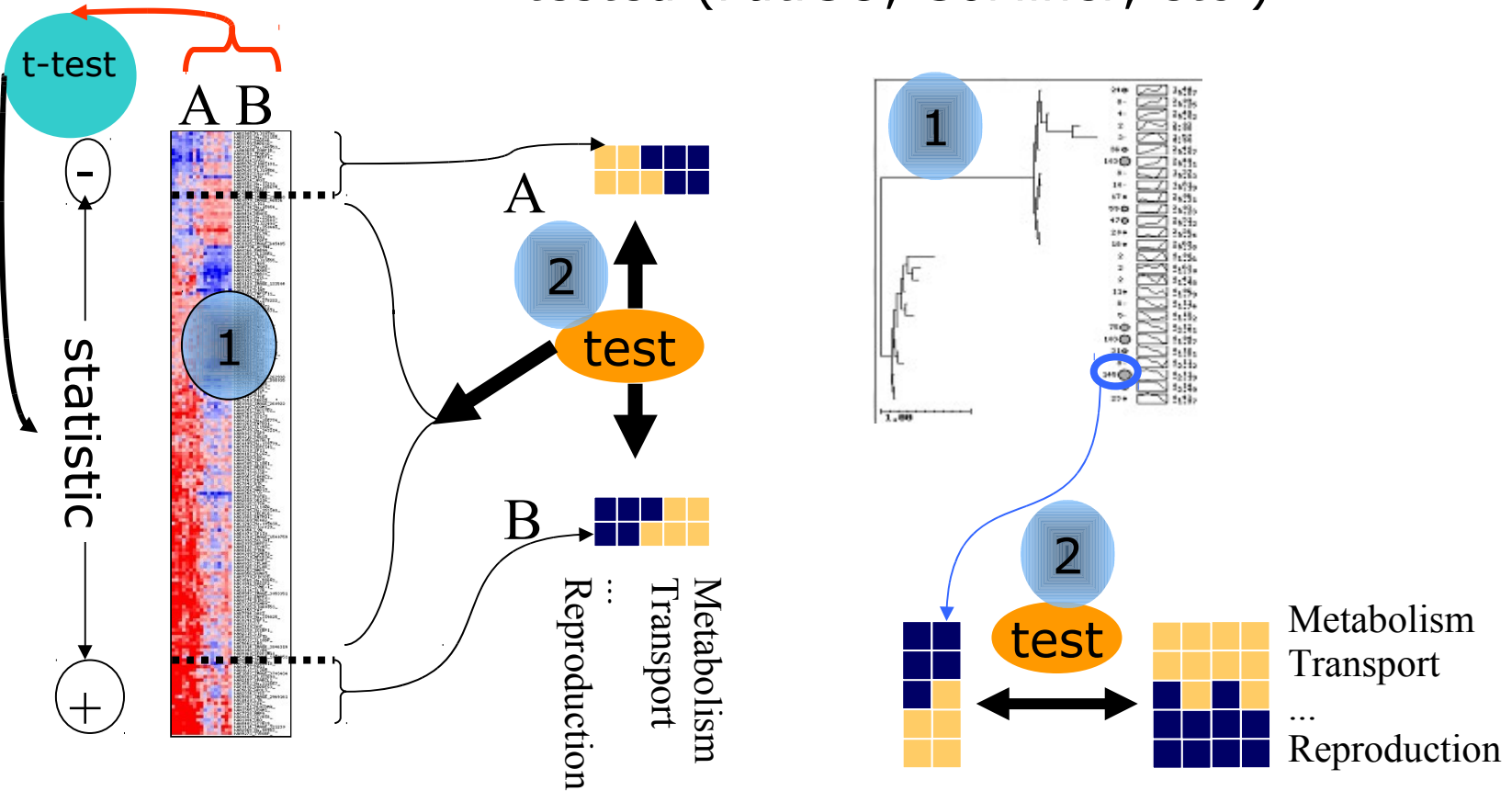
**Regulatory elements**  
miRNA  
CisRed  
Transcription Factor Binding Sites

**Reactome**

**mSigDB**

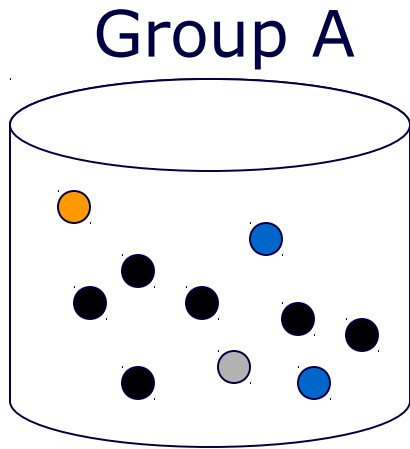
# Two-steps functional interpretation

- 1 Genes are selected based on their experimental values and...
- 2 Enrichment in functional terms is tested (FatiGO, GoMiner, etc.)

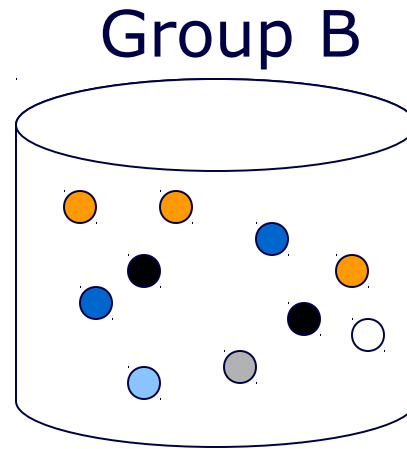


# Testing two GO terms

(remember, we have to test thousands)

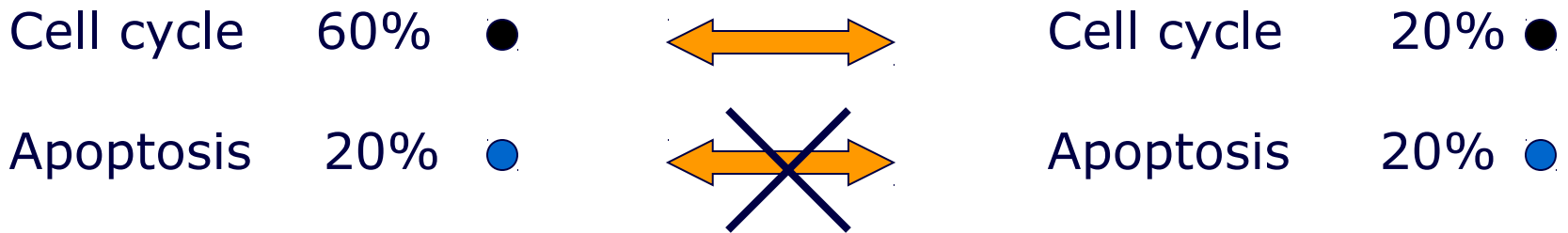


Are this two groups of genes carrying out different biological roles?



	Biosynthesis	Other	
6	4		A
2	8		B

## The popular Fisher's test



Genes in group A have significantly to do with cell cycle, but not with apoptosis.

# GO terms found in sets of 50 genes

GO	Definition	p-value	Adjusted p-value
GO:0006790	sulfur metabolism	0.0595683	1
GO:0042592	homeostasis	0.0157944	0.300094
GO:0016265	death	0.116317	1
GO:0050874	organismal physiological process	0.151987	1
GO:0008152	metabolism	0.129865	1
GO:0019058	viral infectious cycle	0.016503	0.181353
GO:0019059	initiation of viral infection	0.0123062	0.459417
GO:0009056	catabolism	0.0276032	1
GO:0006766	vitamin metabolism	0.00875837	0.604328
GO:0007155	cell adhesion	0.122953	1

Each row corresponds to a random selection of 50 genes from the *E. coli* genome, compared with respect to the rest of the genome.

GO terms in blue (p-value < 0.05 in individual test) have asymmetrical distributions by chance (see adjusted p-values).



# How to test significant differences in the distribution of biological terms between groups of genes?

## FatiGO: GO-driven data analysis

Provides a statistical framework able to deal with multiple-testing hypothesis

The image shows two overlapping browser windows. The left window displays the Gene Ontology (GO) homepage, which includes a navigation menu, a search bar, and a 'Popular Links' section. The right window displays the FatiGO tool page, which provides a detailed description of the tool and its capabilities. An orange arrow points from the 'Tools for using GO' link in the Gene Ontology homepage to the FatiGO tool page.

**Gene Ontology Home**

The Gene Ontology project provides a controlled vocabulary to describe product attributes in any organism. [Read more...](#)

**Popular Links**

Search the Gene Ontology Database

gene or protein name  GO term or ID

This search uses the browser [AmiGO](#). [Browse](#) the Gene Ontology using AmiGO.

**GO website**

- [GO download](#) as including [ontology files](#), [annotations](#) and the [GO database](#).
- [Tools for using GO](#)

**ermineJ** is a tool for the analysis of gene sets (user defined or those defined by GO terms) in expression data. The software is designed to be used by biologists with little or no informatics background. A command-line interface is available for users who wish to script the use of ermineJ. Several different methods for scoring gene sets are implemented, with a focus on methods that don't rely on simple "over-representation" measures.

**FatiGO**

[Bioinformatics Department](#) at the Centro de Investigacion Principe Felipe (Spain) [[PubMed abstract](#)]

[FatiGO](#) assigns representative functional information (under-represented or over-represented Gene Ontology terms) to a given set of genes. Statistical significance is obtained using multiple-testing correction. FatiGO has been designed for functional annotation in the context of DNA microarray data analysis, and is linked to the [Gene Expression Pattern Analysis Suite](#). FatiGO uses gene IDs from the major genomic and proteomic databases (GeneBank, UniProt, Unigene, Ensembl, etc.). FatiGO can also be used for functional annotation of any type of large-scale experiment.

**FuncAssociate**

[Roth Computational Biology Laboratory, Harvard Medical School](#) [[PubMed abstract](#)]

[FuncAssociate](#) is a web-based tool that accepts as input a list of genes, and returns a list of GO attributes that are over- (or under-) represented among the genes in the input list. Only those over- (or under-) representations that are statistically significant, after correcting for multiple hypotheses testing, are reported. Currently 10 organisms are supported. In addition to the input list of genes, users may specify a) whether this list should be regarded as ordered or unordered; b) the universe of genes to be considered by

**Al-Shahrour et al., 2004 Bioinformatics (3rd most cited paper in computing sciences. Source: ISI Web of knowledge.)**

**Al-Shahrour et al., 2005 Bioinformatics. Al-Shahrour et al., 2005 NAR**

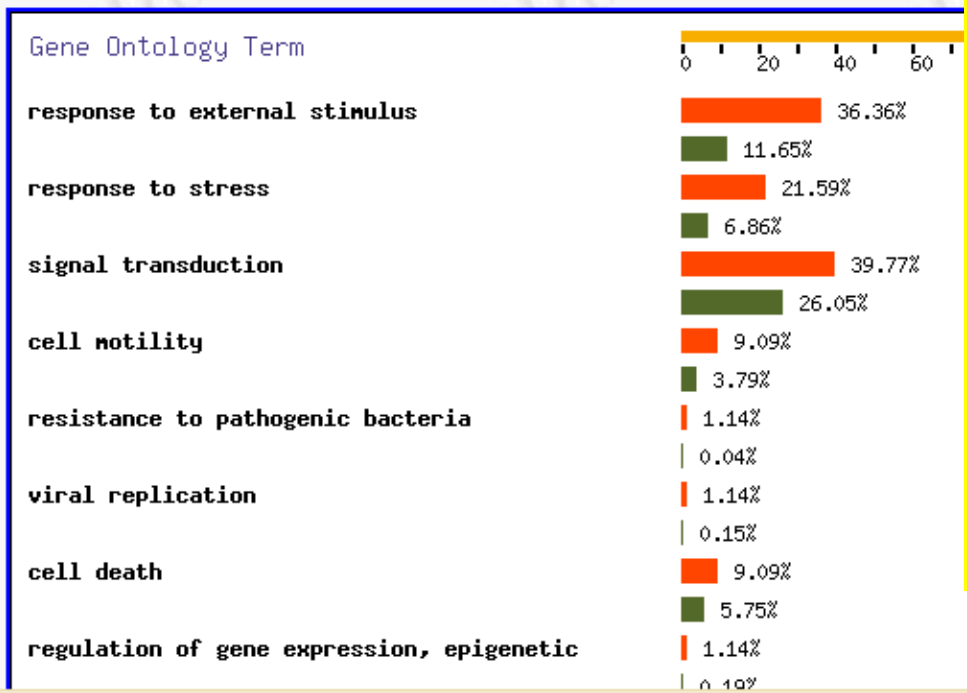
**Al-Shahrour et al., 2006 NAR. Al-Shahrour et al., 2007 BMC Bioinformatics**

**Al-Shahrour et al., 2007 NAR**



# Biological processes shown by the genes differentially expressed among PTL-LB

	Cluster Query	Cluster Reference
Total number of initial genes:	162	4764
Total number of genes no repeated:	129	4731
Total number of Cluster IDs retired - their currents Cluster IDs	7 - 23	449 - 1627
Total number of genes no repeated with current Cluster IDs:	145	5909
Total number of genes no repeated with GO at level 3 and biological_process:	88	2610
Total number of genes no repeated with GO but NOT at level 3 and ontology		
Total number of genes no repeated without GO annotated:		



Obvious? NO

- 1) You now know that there are no other co-variables (e.g. age, sex, etc)
- 2) If you do not have previously a strong biological hypothesis, now you have an explanation

0.1702	0.9912	1	1
0.1806	0.9940	1	1

# Weaknesses of the two-steps, functional enrichment approach

Low sensitivity of conventional gene selection methods

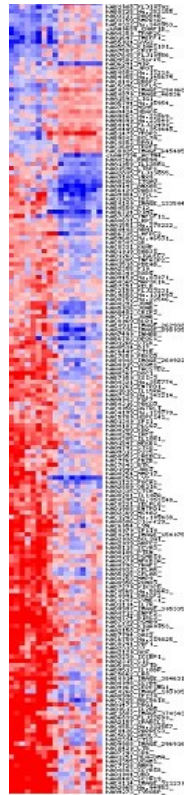
A B

A

8 with impaired tolerance (IGT)  
+ 18 with type 2 diabetes mellitus (DM2)

B

17 with normal tolerance to glucose (NTG)

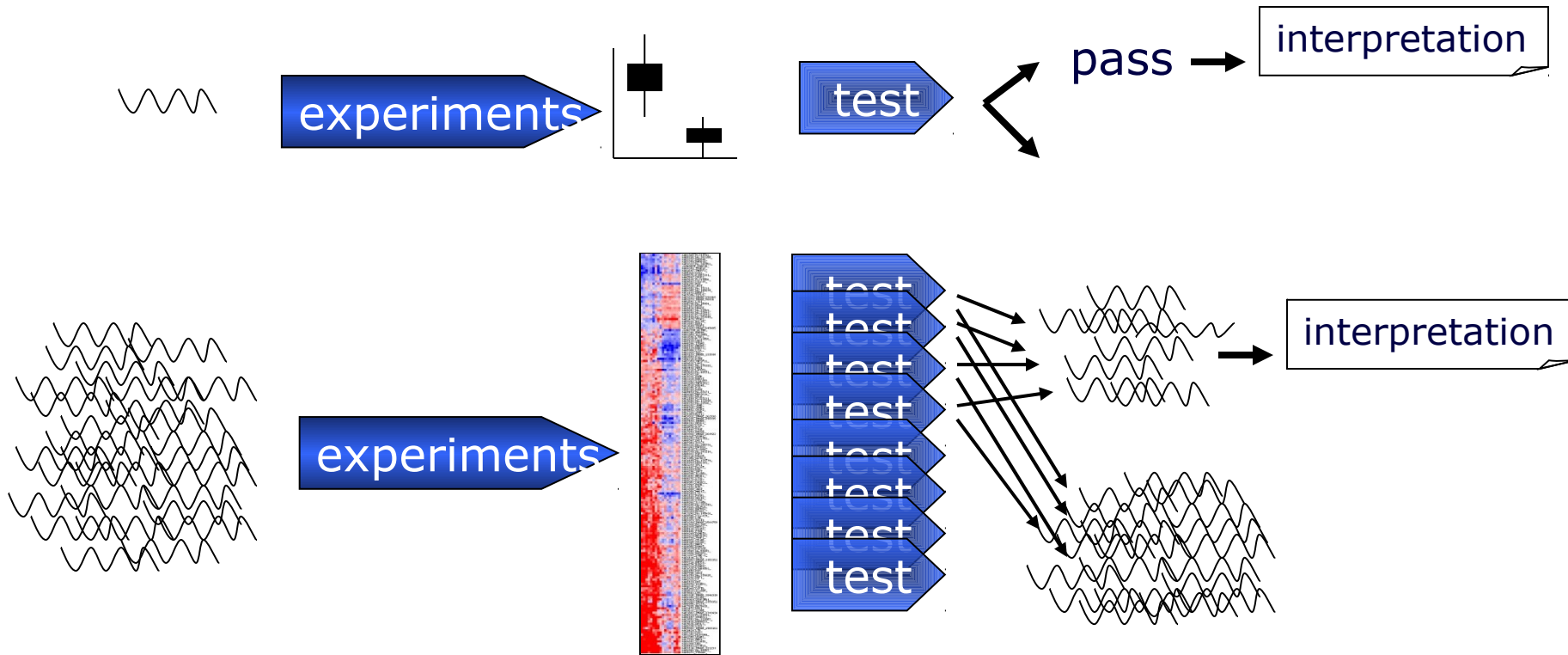


Instability of molecular signatures. Variable selection with microarray data can lead to many solutions that are equally good from the point of view of prediction rates, but that share few common genes (Ein-Dor 2006 PNAS)

Platform comparison. There are still some concerns with the cross-platform coherence of results. Paradoxically, despite the fact that gene-by-gene results are not always the same, the biological themes emerging from the different platforms are increasingly consistent (Bammler 2005 Nat Methods)

*(Mootha et al., 2003)*

# Functional enrichment approach reproduces pre-genomics paradigms



Context and cooperation between genes is ignored

# So, what is wrong with what we are doing?

We seek for the functions activated/deactivated in our experiment

To find them we firstly seek for genes activated/deactivated one at a time (independently)

Then we look among them for enrichment in functions (cooperative activities) using a second test that consider functions independent.

Therefore... is all wrong with this.  
The test we conduct is implicitly answering a question different to the one we want to ask.

# So, what is wrong with what we are doing? (II)

This testing strategy is very strict in controlling:

Type I error ( $\alpha$ ): reject the null hypothesis when the null hypothesis is true, (false positive)

Type II error ( $\beta$ ): fail to reject the null hypothesis when the null hypothesis is false (false negative)

But, we forget about

Type III error : get the right answer having asked the wrong question!

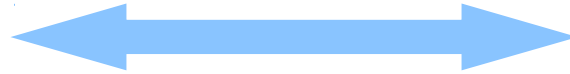
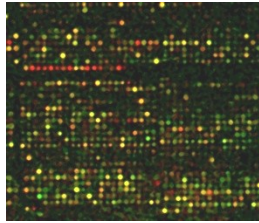
The testing strategy we are conducting is implicitly answering a question different to the one we want to ask.



# Functional genomics.

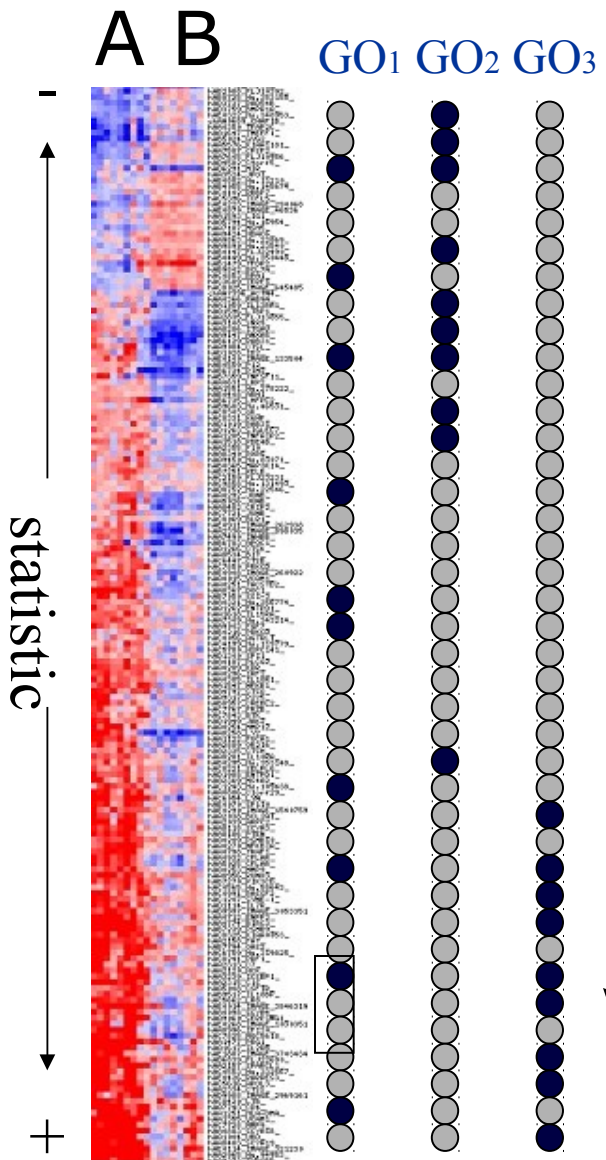
## Historic perspective and future

Differences at phenotype level are the visible cause of differences at molecular level which, in many cases, can be detected by measuring the levels of gene expression. The same holds for different experiments, treatments, strains, etc.



- Classification of phenotypes / experiments. **Sensitivity**
- Selection of differentially expressed genes **Specificity**
- Biological roles the genes are carrying out in the cell. **Interpretation**
- Reformulating the questions. Are we asking the proper questions? What are the real bricks that account for the cellular behaviour and for the phenotype or the response to environmental stimuli? The genes or other higher level units?

# Cooperative activity of genes can be detected and related to a macroscopic observation



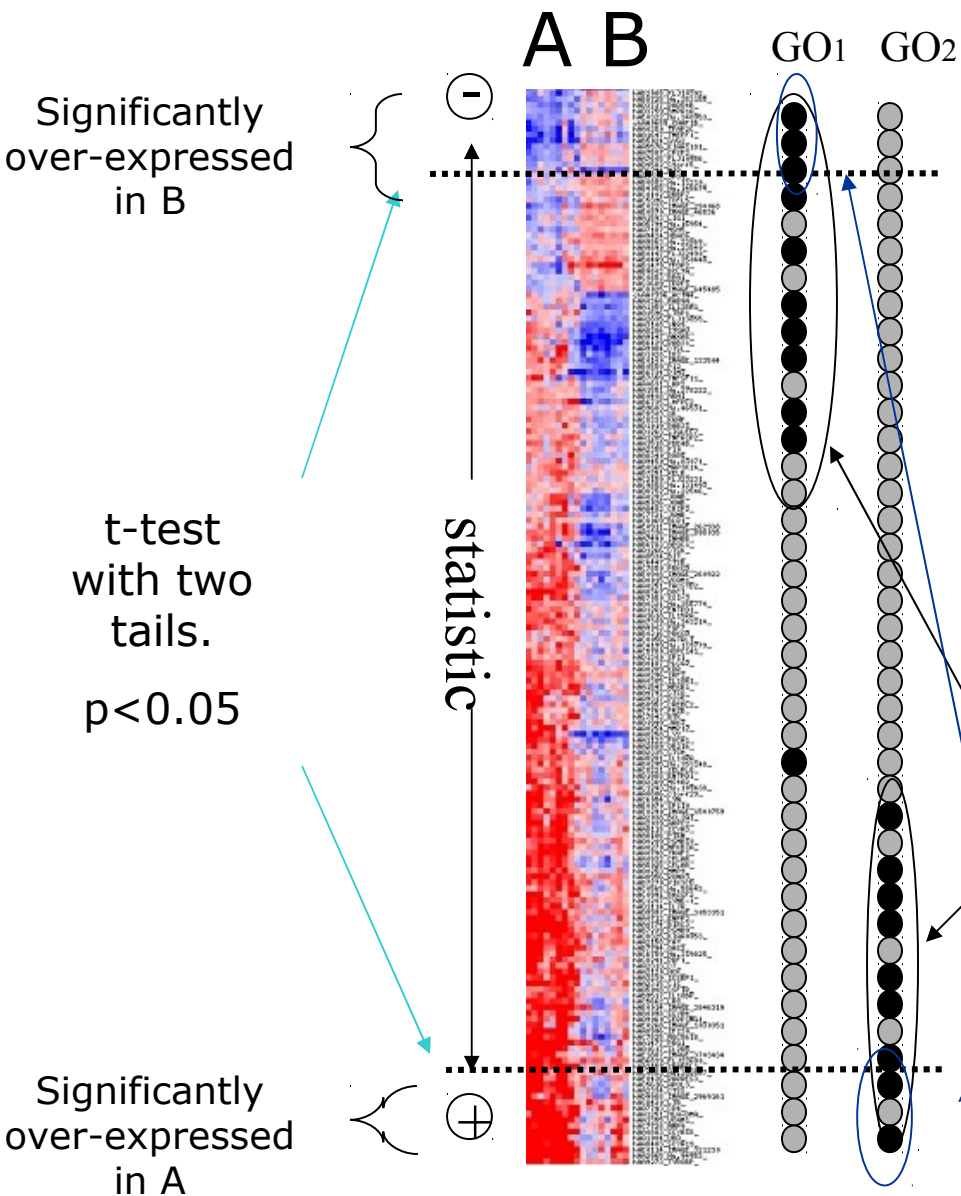
**Ranking:** A list of genes is ranked by their differential expression between two experimental conditions **A** and **B** (using fold change, a t-test, etc.)

**Distribution of GO:** Rows GO1, GO2 and GO3 represent the position of the genes belonging to three different GO terms across the ranking.

The first GO term is completely uncorrelated with the arrangement, while GOs **2** and **3** are clearly associated to high expression in the experimental conditions **B** and **A**, respectively.

Note that genes can be multi-functional

# A previous step of gene selection causes loss of information and makes the test insensitive



If a threshold based on the experimental values is applied, and the resulting selection of genes compared for over-abundance of a functional term, this might not be found.

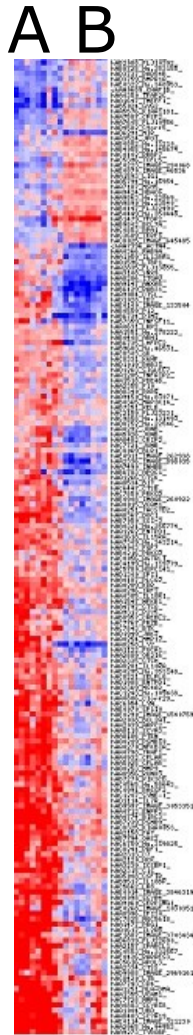
Classes expressed as blocks in A and B

Very few genes selected to arrive to a significant conclusion on GOs 1 and 2

# GSA case study: functional differences in a class comparison experiment

**A**

8 with impaired tolerance (IGT) + 18 with type 2 diabetes mellitus (DM2)



No one single gene shows **significant** differential expression upon the application of a t-test

Healthy vs diabetic	Functional class	GO	KEGG
Up-regulated	Oxidative phosphorylation	X	X
	ATP synthesis		X
	Ribosome		X
	Mitochondrion	X	
	Nucleotide biosynthesis	X	
	NADH dehydrogenase (ubiquinone activity)	X	
	Nuclease activity	X	
Down-regulated	Insulin signalling pathway		X

**B**

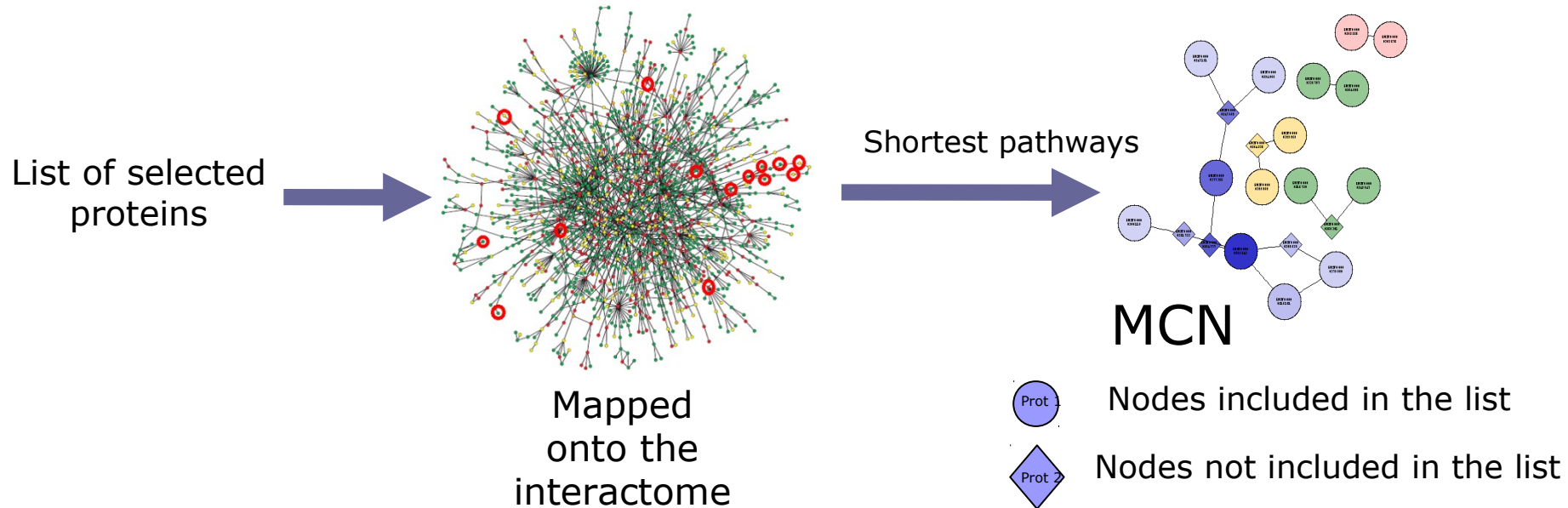
17 with normal tolerance to glucose (NTG)

Nevertheless, many pathways, and functional blocks are **significantly** activated/deactivated

# Protein-protein interaction networks

## Evaluation of the cooperative behaviour of a list of genes

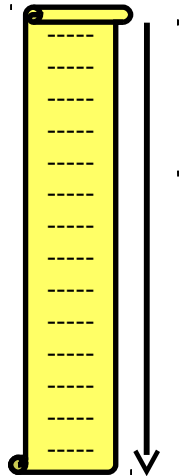
Shortest pathways between all pairs of nodes in the list.  
The minimum connection network (MCN)



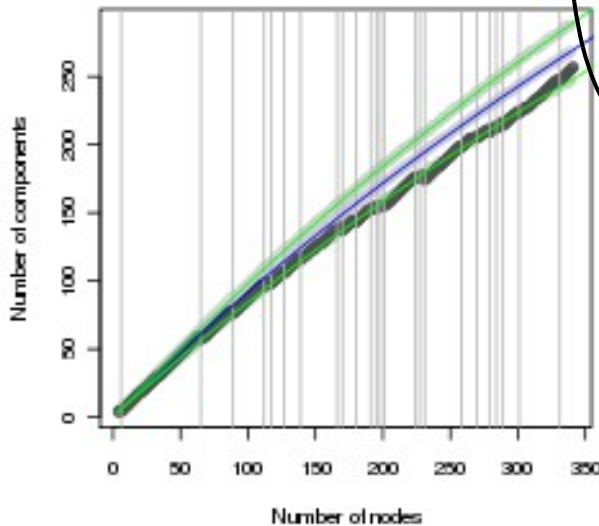


# Gene-set-like network analysis

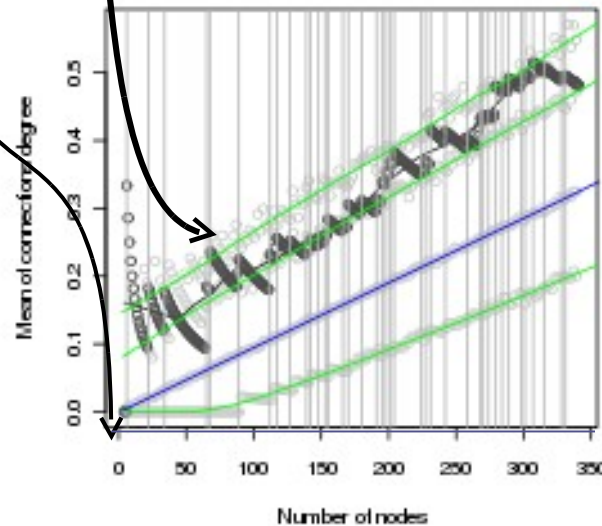
The list is traversed from higher to lower parameter values and the network properties are compared to their random expectations



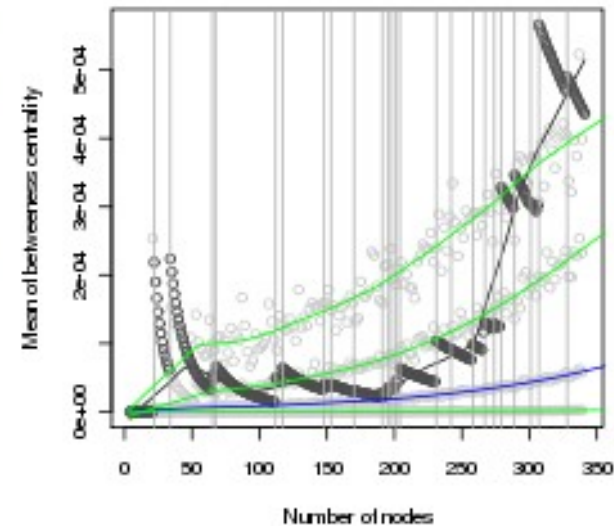
Components REH 0



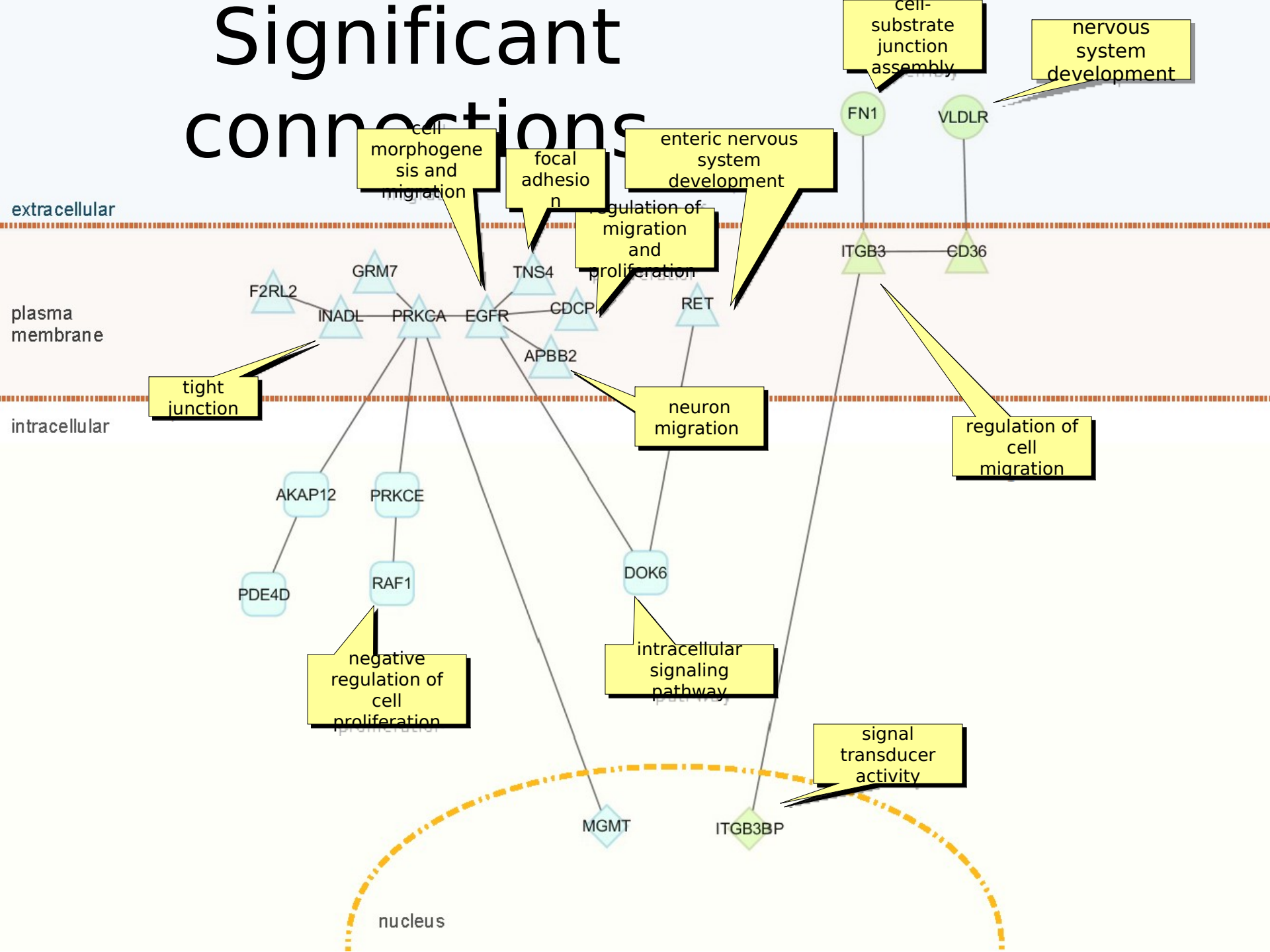
Connections Degree REH 0



Betweenness REH 0



# Significant connections





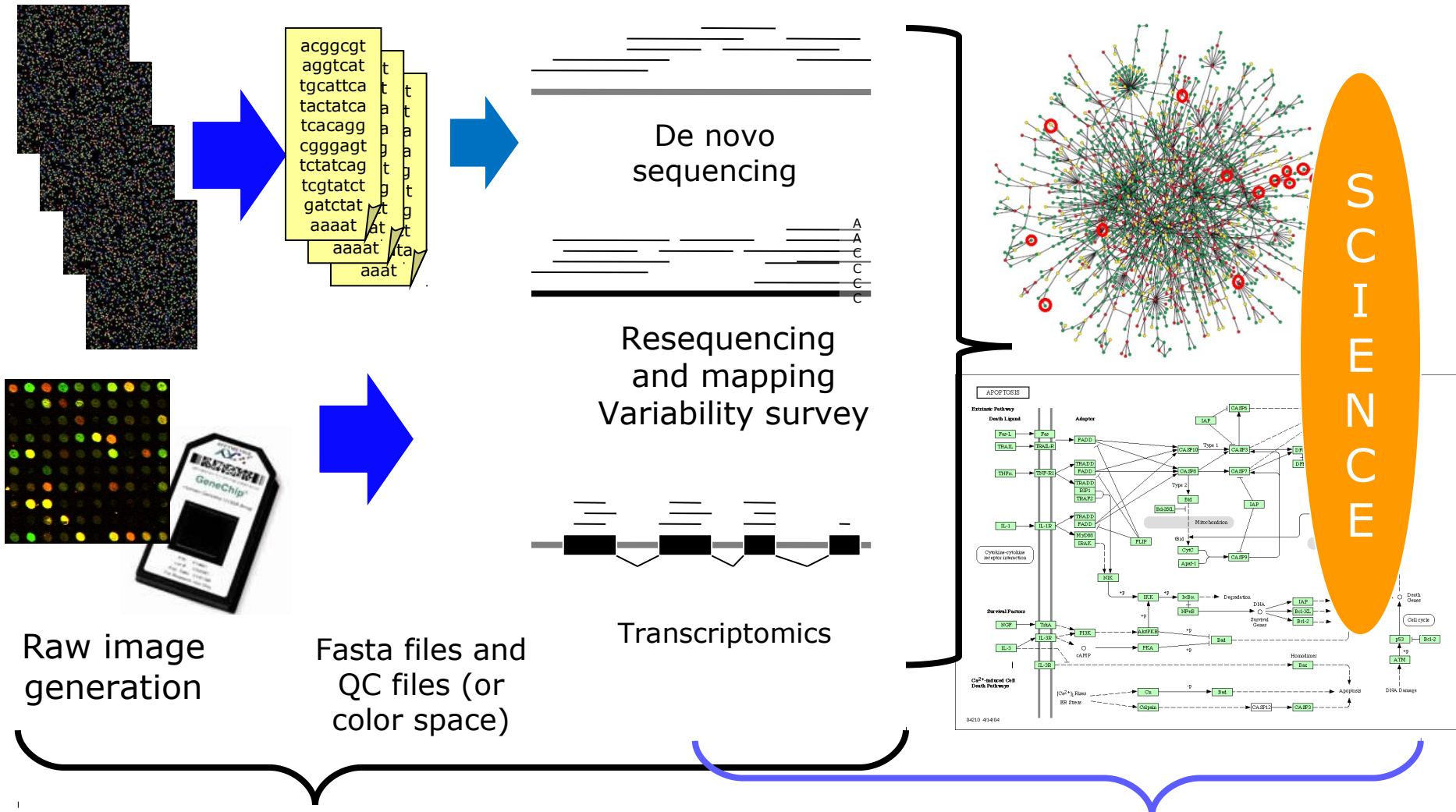
# What is next?

Functional classes have internal structure.  
Exploiting function and internal structure by  
modeling pathways

Method	Gene-based selection	Function-based selection	Function	Relationships among components
Functional enrichment	X		X	
Gene-set analysis		X	X	
Network enrichment	X			X
Network enrichment analysis		X		X
Pathway modeling		X	X	X



# Pipeline general of analysis



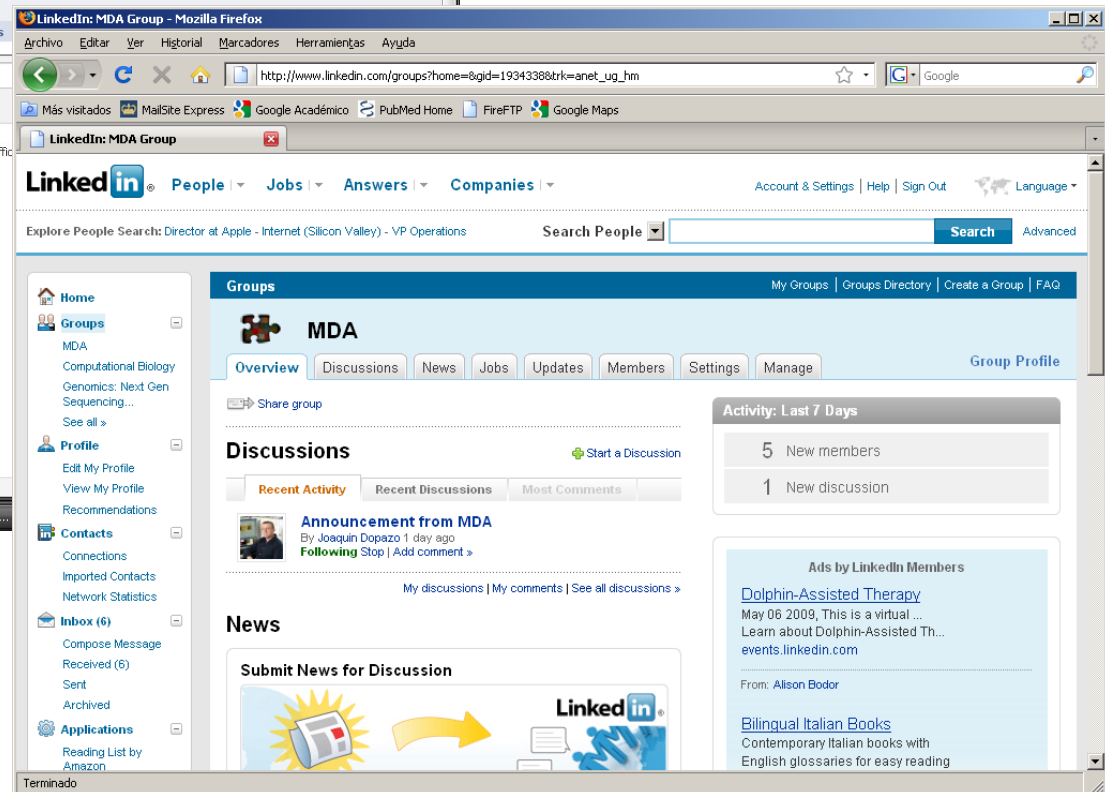
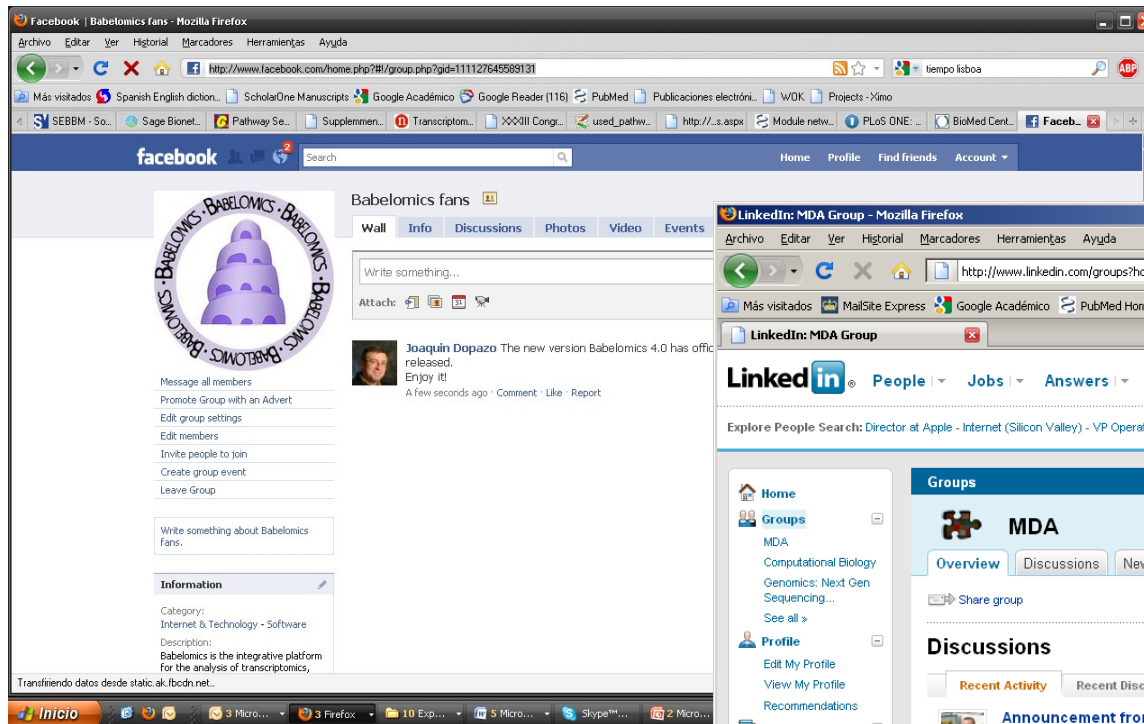
Technology driven

Hypothesis driven

# SOCIAL:

## MDA group in Linked-in

## Babelomics group in Facebook



# The Bioinformatics and Genomics Department at the Centro de Investigación Príncipe Felipe (CIPF), Valencia, Spain, and...



PRINCIPE FELIPE  
CENTRO DE INVESTIGACION



...the INB, National Institute of Bioinformatics  
(Functional Genomics Node) and the CIBERER  
Network of Centers for Rare Diseases, and...

...the Medical Genome Project (Sevilla)

ciberer

