

## Quality Control for RAW Data – Practical session

### 1. Open a terminal window

The programs used in this tutorial are called from the command line. In order to do that, the first step is to open a Terminal window. To do this go to:

**Applications → Accessories → Terminal**



A new window will open with a prompt ready for an input. Now change to the directory with the sequence data. Type on the terminal:

```
cd $HOME/mda13/QC_Raw/
```

### 2. Open FastQC program

To start the FastQC program, you have to type on the terminal window:

```
fastqc &
```

The FastQC application will start in a new window.

You can minimize the terminal window, but **do not close it** while using the FastQC application. Otherwise, FastQC will be also closed.

### 3. Load a file into FastQC

From the FastQC program, go to:

**File → Open**

And load from the folder **\$home/mda13/QC\_Raw** the file called

```
mirna.fastq
```

### 4. Look at the different FastQC result sections and answer the following questions

## Questions:

Sample [mirna.fastq](#)

### 1. Do a quality control for the sample **using FastQC**.

What are the parameters you consider bad quality indicators?

Write down your conclusions:

2. Trim your sample based on its quality with a **minimum quality threshold of 20**.  
You should use `Fastx_toolkit`, so type on the terminal:  
**`fastq_quality_trimmer -t 20 -i mirna.fastq -o mirna_t20.fastq`**

And now you do the quality control for the new file `mirna_t20.fastq` from FastQC.  
What are the main changes?  
Write down your conclusions:

3. Trim the sample based on its quality with a **minimum quality threshold of 28**.  
You should use `Fastx_toolkit`, so type on the terminal:  
**`fastq_quality_trimmer -t 28 -i mirna.fastq -o mirna_t28.fastq`**

And now you do the quality control for the new file `mirna_t28.fastq` from FastQC.  
Is there any quality improvement over the previous filter?  
Write down your conclusions:

4. Trim the sample based on its quality with a **minimum quality threshold of 28**, removing the reads with a **length lower than 30**.  
You should use `Fastx_toolkit`, so type on the terminal:  
**`fastq_quality_trimmer -t 28 -l 30 -i mirna.fastq -o mirna_t28l30.fastq`**

And now you do the quality control for the new file `mirna_t28l30.fastq` from FastQC.  
Is there any quality improvement over the previous step?  
How many reads have been removed?  
Write down your conclusions:

- Trim the sample based on its quality with a **minimum quality threshold of 28**, removing the reads with a **length lower than 35**.

You should use Fastx\_toolkit, so type on the terminal:

```
fastq_quality_trimmer -t 28 -l 35 -i mirna.fastq -o mirna_t28l35.fastq
```

And now you do the quality control for the new file *mirna\_t28l35.fastq* from FastQC.

Is there any quality improvement over the previous step?

How many reads have been removed?

Write down your conclusions:

## Sample **solid.fastq**

1. Do a quality control for the sample using **FastQC**.  
What are the parameters you consider bad quality indicators?  
Write down your conclusions:

2. **Trim** your sample based on its quality with a **minimum quality** threshold of **20**.  
You should use `Fastx_toolkit`, so type on the terminal:  
***fastq\_quality\_trimmer -t 20 -i solid.fastq -o solid\_t20.fastq***

And now you do the quality control for the new file `solid_t20.fastq` from `FastQC`.  
What are the main changes?  
Do you consider the trimming to be effective?  
Write down your conclusions:

3. **Trim** the sample based on its quality with a **minimum quality** threshold of **28**.  
You should use `Fastx_toolkit`, so type on the terminal:  
***fastq\_quality\_trimmer -t 28 -i solid.fastq -o solid\_t28.fastq***

And now you do the quality control for the new file `solid_t28.fastq` from `FastQC`.  
Is there any quality improvement over the previous filter?  
Write down your conclusions:

- Trim the sample based on its quality with a **minimum quality** threshold of **28**, removing the reads with a **length lower than 47**.

You should use `Fastx_toolkit`, so type on the terminal:

```
fastq_quality_trimmer -t 28 -l 47 -i solid.fastq -o solid_t28l47.fastq
```

And now you do the quality control for the new file `solid_t28l47.fastq` from FastQC.

Is there any quality improvement over the previous step?

How many reads have been removed?

Write down your conclusions:

- Remove the reads with **less than a 90%** with **quality above 20**.

You should use `Fastx_toolkit`, so type on the terminal:

```
fastq_quality_filter -q 20 -p 90 -i solid.fastq -o solid_q20p90.fastq
```

And now you do the quality control for the new file `solid_q20p90.fastq` from FastQC.

Has the filter been effective?

How many reads have been removed?

Write down your conclusions:

## Annex 1: Fastx\_toolkit

### fastq\_quality\_trimmer

usage: fastq\_quality\_trimmer [-h] [-v] [-t N] [-l N] [-z] [-i INFILE] [-o OUTFILE]  
Part of FASTX Toolkit 0.0.13 by A. Gordon (gordon@cshl.edu)

- [-h] = This helpful help screen.
- [-t N] = Quality threshold - nucleotides with lower quality will be trimmed (from the end of the sequence).
- [-l N] = Minimum length - sequences shorter than this (after trimming) will be discarded. Default = 0 = no minimum length.
- [-z] = Compress output with GZIP.
- [-i INFILE] = FASTQ input file. default is STDIN.
- [-o OUTFILE] = FASTQ output file. default is STDOUT.
- [-v] = Verbose - report number of sequences.  
If [-o] is specified, report will be printed to STDOUT.  
If [-o] is not specified (and output goes to STDOUT), report will be printed to STDERR.

### fastq\_quality\_filter

usage: fastq\_quality\_filter [-h] [-v] [-q N] [-p N] [-z] [-i INFILE] [-o OUTFILE]  
Part of FASTX Toolkit 0.0.13 by A. Gordon (gordon@cshl.edu)

- [-h] = This helpful help screen.
- [-q N] = Minimum quality score to keep.
- [-p N] = Minimum percent of bases that must have [-q] quality.
- [-z] = Compress output with GZIP.
- [-i INFILE] = FASTA/Q input file. default is STDIN.
- [-o OUTFILE] = FASTA/Q output file. default is STDOUT.
- [-v] = Verbose - report number of sequences.  
If [-o] is specified, report will be printed to STDOUT.  
If [-o] is not specified (and output goes to STDOUT), report will be printed to STDERR.

### Examples:

- Trimming of sequences with quality lower than 20:

```
fastq_quality_trimmer -t 20 -i <sample>.fastq -o <sample_out>.fastq
```

- Trimming of sequences with quality lower than 20 and minimum length of 30:

```
fastq_quality_trimmer -t 20 -l 30 -i <sample>.fastq -o <sample_out>.fastq
```

- Trimming of sequences with less than 90% of bases with quality above 20:

```
fastq_quality_filter -q 20 -p 90 -i <sample>.fastq -o <sample_out>.fastq
```